

The role of vitamin D in the pathogenesis and management of secondary hyperparathyroidism in chronic renal failure

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Summary

Chronic renal failure (CRF) causes alterations in calcium-phosphate metabolism promoting the development of secondary hyperparathyroidism (HPT) and renal osteodystrophy. Recent data suggest that these alterations may play a crucial role in determining cardiovascular calcifications and, thus, cardiovascular mortality among CRF patients. An impaired 1α -hydroxylation of 25-hydroxycholecalciferol (25(OH)D) to 1,25-dihydroxycholecalciferol (1,25(OH)₂D) with decreased circulating 1,25(OH)₂D levels is commonly observed in patients with creatinine clearance below 70 ml/min. The reduction in 1,25(OH)₂D production triggers the upregulation of PTH synthesis, through a decreased suppression on PTH gene transcription and a decreased intestinal calcium absorption. Low vitamin D stores often contribute to the decrease in production and tissue resistance to vitamin D worsens 1,25(OH)₂D deficiency. A reduced expression of vitamin D receptor (VDR) and a less efficient binding of the complex 1,25(OH)₂D-VDR to specific DNA segments account for the resistance to 1,25(OH)₂D in target cells. Thus, absolute and relative 1,25(OH)₂D deficiency is one of the causes of secondary HPT in patients with CRF, together with phosphate retention and skeletal resistance to PTH.

Consistently with these pathophysiological mechanisms, the therapeutic use of 1,25(OH)₂D still represents a milestone for the treatment of uremic secondary HPT and renal osteodystrophy, even though hypercalcemia and hyperphosphatemia are common adverse events and may increase the risk of cardiovascular calcifications. Furthermore, adynamic bone disease may develop after vitamin D therapy. Low levels of serum 25(OH)D are associated with more severe osteodystrophy and HPT, even among dialysis patients. To separate these adverse effects from anti-PTH activity, 1,25(OH)₂D analogues with lower hypercalcemic effect have been synthesized and are now available for clinical use.

KEY WORDS: vitamin D, calcium, parathyroid hormone, phosphate, chronic renal failure.

Vitamin D metabolism

Cholecalciferol (vitamin D₃) and ergocalciferol (vitamin D₂) derive from dietary sources (animal and fish liver, eggs, fish oils). Cholecalciferol is also produced in the skin from 7-dehydrocholesterol (pre-vitamin D₃), through the nonenzymatic effect of sunlight ultraviolet B rays (UVB, wavelengths 295-305 nm). 7-dehydrocholesterol is mostly stored in the cytoplasm of cells at the dermal-epidermal border and the effectiveness of its conversion to cholecalciferol is related to the amount of photochromatic energy entering the skin. Photochromatic energy penetrating the skin greatly depends from the incident angle of UVB rays: in winter, the shallow incident angle of sunlight results in lower energy reaching the epidermis and dermis and, therefore, in lower production of vitamin D₃.

Cholecalciferol, ergocalciferol are hydroxylated at carbon 25 in the liver and at carbon 1 in kidney tubules thanks to enzymatic systems including cytochrome P-450 and located at the inner mitochondrial membrane. PTH and hypophosphatemia enhance the activity of renal 1α -hydroxylase, but not that of liver 25-hydroxylase.

The final product, 1,25-dihydroxycholecalciferol (1,25(OH)₂D), is the active metabolite of vitamin D, although its serum concentrations do not correlate with vitamin D stores. 1,25(OH)₂D promotes active and passive intestinal absorption of calcium and phosphate, and bone mineralization. Conversely, 1,25(OH)₂D suppresses PTH synthesis and parathyroid cell proliferation through a genomic activity. 1,25(OH)₂D₂ and 1,25(OH)₂D₃ have the same potency in activating intestinal calcium absorption and bone mineralization, whereas 1,25(OH)₂D₂ is less potent in suppressing parathyroid gland activity (1). Genomic effect of 1,25(OH)₂D is modulated by specific cytosolic receptors for vitamin D (VDR) in target cells. VDR forms a heterodimer with the retinoid X receptor that enables the complex 1,25(OH)₂D-VDR to bind with high affinity to the vitamin D response element (VDRE) on the transcription promoters of vitamin D-sensitive genes. VDR has been detected in vitamin D-sensitive tissues (bone, intestine, kidney and parathyroid glands) and even in tissues where vitamin D activity is still unclear (myocardium, brain, pancreas and testis). In addition to the genomic effect, a rapid non-genomic effect of 1,25(OH)₂D was found in intestinal cells.

The monohydroxylated metabolite, 25-hydroxycholecalciferol (25(OH)D), is 500 times less active than 1,25(OH)₂D, but its serum concentration is the best indicator of vitamin D body stores. In spite of its low affinity for VDR, 25(OH)D maintains some biological effects, because its serum concentrations are 1000 times higher than those of 1,25(OH)₂D and compensate for the low affinity for VDR (2). The physiopathological relevance of 25(OH)D has been recently reevaluated in population studies showing that low serum concentrations of 25(OH)D were associated with higher serum PTH in healthy elderly individuals (3, 4). In these studies, the serum 25(OH)D concentra-

tion above which all values of serum PTH were normal, was 30 ng/ml (75 nmol/L). This threshold for secondary hyperparathyroidism (HPT) was also confirmed in elderly individuals with elevated creatinine clearance (5) and in hemodialysis patients (6, 7). Based on these findings, the normal range of 25(OH)D serum concentrations was recently redefined and concentration above 30 ng/ml (75 nmol/L) are now generally recommended to prevent secondary HPT (4, 8). Lower 25(OH)D serum concentrations were associated with increased risk of fracture and low bone mineral density (BMD) at different bone sites in young and elderly healthy individuals of both sexes (9, 10). Accordingly, osteopenia and osteoporosis were more frequent in patients with 25(OH)D serum concentrations below 30 ng/ml (75 nmol/L) and osteomalacia was found in patients with 25(OH)D concentrations below 10 ng/ml (25 nmol/L) (4, 8, 11). Serum 25(OH)D concentrations of 30 ng/ml (75 nmol/L) or higher were proposed as the target for the treatment of osteodystrophy, even in hemodialysis patients (6, 7). Hemodialysis patients with lower levels of 25(OH)D had more marked Looser's zone on X-rays and decreased bone formation at bone histology regardless of 1,25(OH)₂D levels (12, 6). However, excessively high 25(OH)D levels were associated with low turnover osteodystrophy. Thus, concentrations between 20 and 40 µg/ml have been proposed as the most appropriate 25(OH)D target range for hemodialysis patients by other authors (12). The need to maintain normal vitamin D stores suggests that unknown vitamin D metabolites, besides 1,25(OH)₂D, may have a beneficial effect on bone and parathyroid metabolism in end stage renal disease.

Vitamin D may have important functions besides mineral ion homeostasis (13). In patients with congestive heart failure, 1,25(OH)₂D concentrations were found to be significantly reduced (14). Recently, it has been demonstrated that vitamin D therapy decreased myocardial hypertrophy in dialysis patients (15). Moreover, vitamin D replacement downregulates the renin-angiotensin system and controls blood pressure in VDR knock-out mice (16). Paradoxically, low 1,25(OH)₂D levels have been correlated with increased coronary calcification in patients at high risk for coronary heart disease (17). Indeed, many epidemiological studies have documented an association between vitamin D deficiency and autoimmune diseases, several types of cancers, and cardiovascular disease (18, 19).

Pathogenesis of secondary hyperparathyroidism in chronic renal failure (CFR)

Parathyroid cells are characterized by a low turnover and rarely undergo mitoses (20, 21). However, in the presence of low calcium, high phosphorus, vitamin D deficiency, and uremia, parathyroid cells leave quiescence and divide by increasing the activity of regulatory cell cycle enzymes and/or their inhibitors (22, 23). In secondary HPT, parathyroid gland growth is initially diffuse and polyclonal. Cell proliferation in the nodules then becomes monoclonal and aggressive (24). The rapid de-differentiation of hyperplastic parathyroid cells in culture (25) precludes further assessment of the relative contribution of changes in calcium, phosphate, and vitamin D to the expression of components of the cell cycle critical for growth control.

Role of phosphate

Hyperphosphatemia due to decreased glomerular filtration rate is an important factor in the pathogenesis of secondary HPT (26). Elevated serum phosphate levels induce secondary HPT through indirect and direct mechanisms (27). In addition, hyperphosphatemia inhibits 1,25(OH)₂D production (28), with subsequent hypocalcemia.

The direct effects of phosphorus have been demonstrated both *in vitro* and *in vivo* studies. High phosphorus concentrations stimulate PTH secretion in intact rat parathyroid glands (29). Unfortunately, the *in vitro* effects of phosphorus on PTH secretion could be observed in intact parathyroid tissue preparations, but not isolated, dispersed parathyroid cells (12, 30). Recently, several studies have shown that phosphate may regulate parathyroid function at post-transcriptional level, as it improves PTH mRNA stability through binding of parathyroid cytosolic proteins to the 3'-UTR and especially to the terminal 60 nucleotides of PTH mRNA (31).

Two weeks after 5/6 nephrectomy in rats, uremia-induced mitotic activity is further enhanced by high dietary phosphate, but prevented by phosphorus restriction (22). In contrast to the mitogenic effects of hyperphosphatemia, dietary phosphorus restriction appears to counteract the proliferative signals induced by uremia, thus preventing parathyroid cell replication and the increase in parathyroid gland size (32).

Table I - Values of the parameters of calcium-phosphate metabolism in patients with different creatinine clearance. A significant decrease of serum 1,25(OH)₂D is observed since creatinine clearance values of 70 mL/min and is associated with secondary HPT. Serum phosphate increases later, but occurs in the early stages of CRF. Intestinal calcium absorption decreases during the progression of CRF. It was measured using strontium (Sr) as a marker: strontium absorption was determined 4 hours after an oral load and expressed as area under the time-plasma strontium concentration curve (90).

N.	9	10	11	10	18
Creatinine clearance (mL/min)	<20	21-40	41-60	61-80	>80
Age (years)	56±4.7	56±3.4	59±2.6	50±5.5	52±2.1
Serum phosphate (mmol/L)	1.8±0.09	1.26±0.07	1.17±0.05	1.12±0.05	1.1±0.04
Plasma calcium (mmol/L)	2.08±0.1	2.38±0.03	2.42±0.03	2.35±0.03	2.35±0.02
Serum 1,25(OH) ₂ D	15±2.9	21±3.2	23±3.6	28±2.9	39±2.8
iPTH (pg/ml)	494±120	129±37	47±7	63±12	29±3
Enteral Ca absorption (mmol of absorbed Sr/l per min)	3.4±0.4	6.2±0.9	6.1±0.4	7.6±0.7	8.1±0.6

Recent studies on the effects of dietary phosphate on parathyroid gland growth demonstrated that the low phosphorus-induced of p21 (an inhibitor of kinases of the cell-cycle) mRNA and protein content in parathyroid glands contributes to the antiproliferative effects of phosphate restriction on uremia-induced parathyroid cell growth (33). In this uremic rat model, the temporal increases in p21 protein expression correlate inversely with the parathyroid levels of a marker of mitotic activity, the proliferating nuclear cell antigen (PCNA) (33).

In the search for the mitogenic stimuli triggered by high dietary phosphate, we next focused on transforming growth factor- α (TGF α). TGF α , known to promote growth not only in malignant transformation but also in normal tissues (34), is increased in hyperplastic and adenomatous human parathyroid glands (35). High phosphate diet worsens uremia-induced parathyroid hyperplasia by increasing parathyroid expression of TGF α . The rapid return of parathyroid TGF α content to normal after phosphate restriction suggests that low phosphorus may counteract uremia-induced parathyroid cell growth not only through induction of p21 expression, but also by preventing the increase in parathyroid TGF α (33, 36).

Increase in parathyroid TGF α induces cell growth through autocrine and paracrine mechanisms through activation of its receptor, the epidermal growth factor receptor (EGFR) (37). In human parathyroid glands, Gogusev et al. (35) demonstrated the presence of EGFR protein in 4 out of 5 adenomas, in 13 out of 15 tissue samples of hyperplasia secondary to renal failure, and in most samples of normal parathyroid tissue. No differences in the expression patterns were observed between groups. However, studies in 104 human hyperplastic parathyroid glands, which failed to detect EGFR protein, showed higher EGFR mRNA expression in carcinoma and primary hyperplasia compared to adenomas and hyperplasia secondary to renal failure (38).

The concept that co-expression of TGF α and EGFR could contribute to non-neoplastic parathyroid hyperplasia led us to examine the dietary-phosphate regulation of parathyroid EGFR expression in rat parathyroid glands. Similarly to the changes in TGF α expression, high dietary phosphate increases parathyroid EGFR content to above normal levels, while phosphorus restriction prevented the increases in EGFR levels (39, 40).

These findings indicate that the uremia-induced parathyroid co-expression of TGF α and its receptor, EGFR, acts as a mitogenic signal, which can be blunted by phosphate restriction and counteracted through the induction of p21. These new insights into the molecular mechanisms of parathyroid hyperplasia suggest that, in addition to phosphate restriction or use of phosphate binders, therapeutic approaches focusing on induction of p21 and inactivation of TGF α /EGFR growth-promoting signals may slow down the progression of secondary HPT.

Role of calcium

Calcium is a key regulator factor in secondary HPT progression. Low serum calcium levels decrease the activation of the Calcium-Sensing Receptor (CaSR), a plasma membrane G-protein coupled molecule that allows parathyroid cells to sense calcium in the extracellular fluid, thus greatly promoting PTH synthesis and secretion (41). In contrast, hypercalcemia activates the CaSR, rapidly suppressing secondary HPT. Recent evidence suggests that signaling through the CaSR plays an important role on parathyroid hyperplasia (42). Moreover, calcium-dependent signaling through the CaSR may prevent parathyroid hyperplasia even in tissues that are not-responders to vitamin D (43).

The extracellular calcium concentration may also regulate the level of PTH mRNA (44, 45) and parathyroid cell proliferation (46). Moreover, calcium regulates VDR mRNA and protein ex-

pression in parathyroid cells independently of 1,25(OH)₂D (47). Clearly, serum calcium levels could also indirectly regulate PTH levels through a feedback of 1,25(OH)₂D on the parathyroid glands.

Reduced expression of CaSR in hyperplastic parathyroid glands has been demonstrated in an animal model (48). Parathyroid hyperplasia appears 48 hours after 5/6 nephrectomy in rats fed a high phosphate diet, while CaSR expression starts to decrease 48 hours later. Furthermore, in humans, CaSR content declines by about 60% in hyperplastic parathyroid glands compared to normal controls (49).

Further support for the pathophysiological relevance of changes in the expression of parathyroid p21, TGF α and EGFR in controlling proliferative activity came from studies, evaluating the expression of these three proteins after suppression of parathyroid cell growth by high-calcium intake or its further enhancement by low dietary calcium. High dietary calcium controlled uremia-induced parathyroid hyperplasia, reducing both parathyroid gland size and the expression of two markers of mitotic activity, Ki67 and PCNA (36). Furthermore, high calcium diet increased parathyroid p21 levels and prevented the rise in parathyroid content of TGF α and EGFR induced by uremia (36, 40). The mechanisms for high-calcium induction of p21 and prevention of the increase in TGF α and EGFR are unknown. Studies in vitamin D receptor-ablated mice showed the ability of a calcium-enriched diet to prevent the development of parathyroid hyperplasia in both hypocalcemic and normocalcemic states (50).

In relation to high-calcium control of TGF α /EGFR growth promoting signal, hypercalcemia and low plasma levels of TGF α were recently associated in cancer patients, suggesting the possibility of the systemic control of TGF α expression by calcium (51). The existence of such an association in CRF patients could partially explain the suppression of parathyroid growth by hypercalcemia.

The changes in p21, TGF α and EGFR in the parathyroid glands of uremic rats fed a high Ca diet suggest that increases in serum calcium or in intracellular calcium, induced by vitamin D therapy, could enhance the effects of vitamin D itself in increasing p21 expression and reducing TGF α and EGFR content.

Role of vitamin D

In the 5/6 nephrectomized rats, 1,25(OH)₂D₃ suppresses uremia-induced parathyroid cell proliferation both *in vitro* (52), and *in vivo* (53).

Naveh-Many showed that PTH mRNA was much higher in parathyroid glands from vitamin D-deficient normocalcemic rats than controls, and that in vitamin D-deficient hypocalcemic rats the upregulation of PTH mRNA was even more pronounced (54). Moreover, many studies have been conducted to assess whether supplementation with vitamin D sterols can prevent or ameliorate secondary HPT in CRF.

In the early-uremia rat model (7 days of renal failure), 1,25(OH)₂D and the less hypercalcemic vitamin D analog 1,25-dihydroxy-19-norvitamin D₂ (19-norD₂) controlled both serum PTH levels and parathyroid hyperplasia similarly to what is described with phosphate restriction (36). The suppression of uremic rat parathyroid cell growth by vitamin D treatment can be partially accounted for by the increased expression of p21. Furthermore, studies in patients with secondary HPT suggest an important role for increased p21 expression in parathyroid growth arrest (55).

The efficacy of either 1,25(OH)₂D or 19-norD₂ to arrest parathyroid hyperplasia and parathyroid gland enlargement, was associated with prevention of TGF α and EGFR expression in the

parathyroids (39). Since the TGF α activation of its receptor induces both TGF α and EGFR gene expression, it is also possible that 1,25(OH) $_2$ D inhibition of EGFR activation mediates the suppressive effects of the sterol on TGF α and EGFR expression (56).

In addition, vitamin D therapy suppresses parathyroid growth even in an established rat model of secondary HPT. 5/6 nephrectomized rats fed a high phosphorus diet for 8 weeks and received either vehicle or the vitamin D analog 19-norD $_2$. No increase in parathyroid gland size was observed in the vitamin D treated uremic animals compared to the untreated rats (36).

In patients with CRF (Table I), the inability to synthesize 1,25(OH) $_2$ D is sustained by decreased 1 α -hydroxylation of 25(OH)D in tubular cells and occurs prior to the increase in PTH secretion (57). Serum concentrations of 1,25(OH) $_2$ D start to decrease at values of creatinine clearance near 70 mL/min. Therefore, patients are predisposed to secondary HPT even in the early phases of CRF (58). Because vitamin D sensitises the parathyroid gland to calcium, it is possible that vitamin D deficiency in early CRF may contribute to the development of secondary HPT, even in the absence of overt hypocalcemia.

Alteration of renal 1 α -hydroxylase is frequently associated with other defects that may further diminish 1,25(OH) $_2$ D synthesis. A reduction in vitamin D stores may be found in a significant proportion of CRF patients, but also of healthy individuals in industrial countries (3, 59). Among healthy elderly Europeans 36% of men and 47% of women had serum 25(OH)D levels below 12 ng/mL (30 nmol/L), with the highest prevalence in Southern countries (60, 61). Similar frequencies were also observed in non-European populations (62, 11). Ninety-three percent of Spanish elderly individuals with plasma creatinine higher than 1 mg/dL, and 79% of Italian patients with creatinine clearance between 30 and 70 mL/min, had serum 25(OH)D below 30 ng/ml (75 nmol/L) (5, 63). In these populations, 25(OH)D deficiency was not necessarily associated with a decrease in serum concentrations of 1,25(OH) $_2$ D (5). Inactivity with insufficient exposure to sunlight or low dietary intake may be responsible for vitamin D insufficiency in CRF patients. Patients with CRF may also develop a reduced capability to produce 25(OH)D and resistance to UVB, due to skin hyperpigmentation caused by retention of melanocyte-stimulating hormone. An additional factor may be patient's age, since aging is usually associated with lower skin concentration of 7-dehydrocholesterol and decreased responsiveness of renal 1 α -hydroxylase to PTH (64). Underlying renal disease may also affect vitamin D status. Urinary loss of vitamin D bound to vitamin D-binding protein occurs in nephrotic syndrome. Likewise, patients undergoing peritoneal dialysis lose vitamin D-binding protein with peritoneal effluent.

1,25(OH) $_2$ D concentrations were directly related with those of 25(OH)D in CRF patients (62), but not in healthy individuals. Thus, the activity of renal 1 α -hydroxylase in CRF appears to be substrate-dependent and is depressed by low serum concentrations of 25(OH)D (63, 62, 65). Interestingly, a positive correlation between 25(OH)D and 1,25(OH) $_2$ D was also observed in hypercalciuric stone formers with normal creatinine clearance (66). Therefore, careful assessment of 25(OH)D status is today proposed as an integral part of the therapeutic strategy to prevent secondary HPT and bone disease in CRF patients.

Clinical consequences of secondary hyperparathyroidism

Secondary HPT causes both skeletal and extra-skeletal complications. Bone-associated consequences include with osteitis

fibrosa and renal osteodystrophy, while non-skeletal consequences include cardiovascular calcification, soft-tissue calcification, endocrine disturbances, compromised immune system, neurobehavioral changes, and altered erythropoiesis (67, 68).

Renal osteodystrophy

Renal osteodystrophy refers to a bone disorder that occurs in patients with CRF, characterized by abnormal bone turnover (69). PTH directly stimulates calcium mobilization from the bone by acting on the osteoclasts and indirectly by stimulating bone resorption and formation through the increase of vitamin D synthesis in the kidney. It is well known that vitamin D stimulates osteoblast activity and increases both osteoclast amount and function (69). In CRF patients vitamin D deficit leads to a loss in normal osteoblast activity and a reduction in bone mineralization. Furthermore, high serum PTH levels stimulate osteoclasts to mobilize more calcium from bone tissue, with bone mass reduction (70, 71).

In CRF patients renal osteodystrophy is characterized by five types of bone lesions. Osteitis fibrosa and mixed osteodystrophy are high-turnover bone diseases. Osteomalacia, aluminium-induced bone disease, and adynamic bone disease are low-turnover bone diseases (69). High-turnover bone diseases are characterized by rapid bone turnover induced by abnormally high serum PTH levels and abnormal mineralization. Conversely, low-turnover bone diseases are caused by over-suppression of PTH or aluminium at the site of bone mineralization (69). Figure 1 displays the complex interaction among the multiple factors that may cause the different bone lesions in renal osteodystrophy.

Cardiovascular calcification

CRF patients have a dramatically higher incidence of cardiovascular morbidity and mortality compared to the general population. In the last ten years, several studies pointed out that vascular calcification is a major cause of cardiovascular disease in the dialysis population. In CRF patients, high levels of plasma calcium, serum phosphate and PTH play a critical role

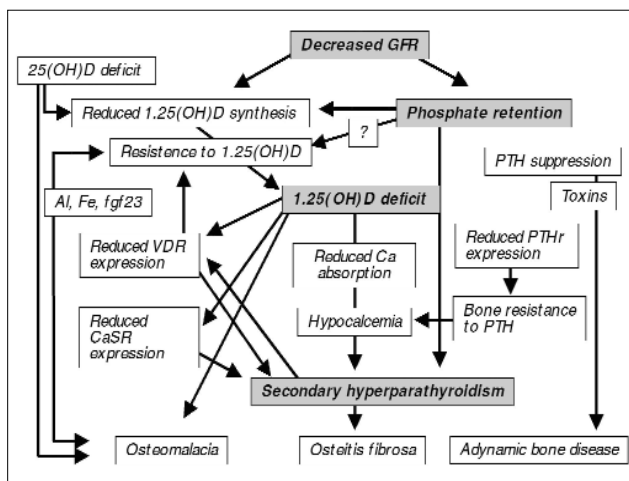


Figure 1 - This figure displays the complex interaction between the different components of the system controlling calcium and phosphate in blood. The decrease of glomerular filtration rate (GFR) leads to phosphate retention and 1,25(OH) $_2$ D deficiency that induce parathyroid hyperplasia and stimulate PTH secretion. Various endogenous or exogenous toxins interfere with bone mineralization processes.

in the pathogenesis of cardiovascular events.

Patients develop extensive medial calcification, which causes increased arterial stiffness and high morbidity and mortality due to cardiovascular events (72, 73). A variety of risk factors are associated with vascular calcification in dialysis patients (time on dialysis, uremic toxins, history of diabetes, inflammation), but abnormalities in bone mineral metabolism may play a critical role (74). In fact, elevated serum phosphate, calcium-phosphate product, and PTH levels contribute to vascular calcification, although their roles are incompletely understood (75, 76).

Elevations of serum phosphate and calcium-phosphate product levels may worsen cardiovascular events in the uremic population, through a progressive increase in calcium deposition in the coronary arteries and heart valves (77). Furthermore, calcium-containing phosphate binders can increase calcium load (78). Recently, an elegant study by Goodman et al. (79) in hemodialysis children and young adults showed a correlation between coronary artery calcification detected by electron beam computed tomography (EBCT) and age of dialysis, serum phosphorus, calcium-phosphate product levels, and daily intake of calcium. Moreover, experimental observations in uremic rats (80, 81) and in dialysis patients (82) showed that a new calcium and aluminium-free phosphate binder (Sevelamer HCl) reduced the progression of cardiovascular calcification more than calcium-based phosphate binders.

Treatment of secondary hyperparathyroidism: role of vitamin D and its analogues

Prevention and treatment of secondary HPT commonly requires control of both phosphate and $1,25(\text{OH})_2\text{D}$ levels in serum. Phosphate levels are usually controlled by reducing dietary phosphate intake, by dialysis, and by using phosphate-binders. $1,25(\text{OH})_2\text{D}$ effectively suppresses PTH production and improves bone histology in some patients (77). Vitamin D and its analogues act after binding to VDR in parathyroid cell cytosol. The vitamin D-VDR complex controls the rate of PTH gene transcription by interacting with VDRE, located 100-120 bases upstream from the PTH gene (83). Furthermore, vitamin D upregulates CaSR expression on cell membrane, thus leading to a greater inhibition on PTH secretion (41). Through CaSR the increase in circulating calcium ions can inhibit tonic PTH secretion by stimulation of mitogen-activated protein kinase pathways (84). VDRE regions are present in the two promoters of the CaSR gene (85). Calcium ions inhibit PTH secretion not only through CaSR, but also by interacting with cytosolic proteins affecting the stability of PTH mRNA. Hypocalcemia favors the activity of cytosolic proteins protecting PTH mRNA from the degrading effect of ribonucleases. On the other hand, hypercalcemia favors the activity of proteins degrading PTH mRNA (86). Furthermore, a transcriptional activity of calcium has been hypothesized (87). Posttranscriptional and transcriptional effects of calcium ions are indirectly influenced by the calcium-repleting activity of vitamin D.

The therapeutic use of $1,25(\text{OH})_2\text{D}$ in CRF mainly aims to raise intestinal absorption of calcium, to protect bone against osteomalacia and to control parathyroid function. However, also the function of other organs expressing VDR, as kidney, pancreas, myocardium and testis, might be improved by vitamin D administration. $1,25(\text{OH})_2\text{D}$ inhibitory effect on PTH synthesis and parathyroid hyperplasia is well established in CRF. Vitamin D analogues with a lower hypercalcemic and hyperphosphatemic activity, are now available as an efficient alternative to $1,25(\text{OH})_2\text{D}$ treatment. Furthermore, normal serum levels of $25(\text{OH})\text{D}$ are relevant for the prevention or treatment of sec-

Table II - Objectives for renal osteodystrophy treatment in patients with different levels of CRF recommended in K/DOQI guidelines. Stage 1 and 2 of renal function are characterized by creatinine clearance above 90 mL/min and between 60 and 90 mL/min.

	Stage of renal function		
	Stage 5	Stage 4	Stage 3
Creatinine clearance (mL/min)	<15	16-30	30-60
iPTH (pg/mL)	150-300	70-110	35-70
Serum phosphate (mmol/L)	1.13-1.78	0.87-1.49	0.87-1.49
Plasma calcium (mmol/L)	2.1-2.37	2.2-2.5	2.2-2.5
Ca-P product (mmol ² /L ²)	< 4.5	< 4.5	< 4.5

ondary HPT, due to the capacity of $25(\text{OH})\text{D}$ to support renal 1α -hydroxylase activity or the synthesis of active vitamin D metabolites different from $1,25(\text{OH})_2\text{D}$ (6, 62, 63, 65).

Clinical targets for serum concentrations of phosphate, calcium and PTH (Table II) have been proposed in the 2003 guidelines proposed by the Kidney Dialysis Outcomes Quality Initiative (K/DOQI) Work Group (88). Target for plasma concentrations of PTH increases according to creatinine clearance because patients develop a progressive tissue resistance to PTH explained by a diminished expression of PTH receptors and by $1,25(\text{OH})_2\text{D}$ deficiency or resistance (89, 90). The low target for calcium concentrations aims to avoid calcium overload and an excessively high calcium-phosphate product.

A particular bone complication of the therapy with vitamin D metabolites occurs in case of excessive parathyroid suppression. As defined in the K/DOQI guidelines, the target for PTH concentration is 150-300 pg/mL in end-stage CRF (Table II). If the values of circulating PTH are suppressed below these limits, bone cells become quiescent and bone remodeling is far slowed (91). The histology pattern of these functional alterations is adynamic bone disease, characterized by decreased number of bone cells, bone metabolic inactivity and reduced bone formation rate (91). The mechanism of adynamic bone disease is unknown. The clinical consequences of adynamic bone disease may be an increased fracture risk, while its relationship with osteoporosis is still debated (92). At the opposite, osteomalacia may develop in case of $1,25(\text{OH})_2\text{D}$ deficiency, even though the toxic effect of aluminum overload has been the most frequent cause of osteomalacia in dialysis population till some years ago (93).

Vitamin D supplementation

Vitamin D deficit is frequent in CRF and its correction prevents or cures secondary HPT and achieves the specific beneficial effects of $25(\text{OH})\text{D}$ on muscles and bones (6, 94). Vitamin D supplements are recommended in patients with creatinine clearance above 15 mL/min and with serum concentrations of $25(\text{OH})\text{D}$ below 30 ng/mL (75 nmol/L), because their kidney is stimulated to produce $1,25(\text{OH})_2\text{D}$ by the increase of 1α -hydroxylase substrate, $25(\text{OH})\text{D}$. Ergocalciferol is the most convenient product to replace vitamin D stores, due to its availability, cost and efficacy. K/DOQI guidelines recommend to administer 50000 IU/month per six months orally in patients with serum $25(\text{OH})\text{D}$ concentrations of 16-30 ng/mL (40-75 nmol/L), or six-month courses with more frequent doses in the presence of lower $25(\text{OH})\text{D}$ levels (88). During this treatment, plasma

calcium concentration should not exceed 2.51 mmol/L (10.5 mg/dL) and serum phosphate 1.49 mmol/L (4.6 mg/dL). To prevent vitamin D deficiency in CRF patients, Schomig and Ritz proposed oral supplementation with 1000 IU/day of cholecalciferol, or 10000 IU/week (58). Alternatively, 25(OH)D may be administered orally at the dose of 10-40 µg/day, according to the serum levels of this metabolite and calcium, as usually employed in osteoporosis.

The use of vitamin D in end-stage CRF has been source of debate. Hemodialysis patients with low levels of 25(OH)D had more marked Looser's zones at the pelvis radiograms and decreased bone formation at bone histology, regardless of 1,25(OH)₂D levels (12, 6). Gazali et al. suggest to prescribe 1600 IU/day of vitamin D or 20-30 µg/day of 25(OH)D at least during the winter months (6). The doses of vitamin D proposed by these Authors were higher than those commonly indicated for general elderly population (800 IU/day), because the need of vitamin D is increased in CRF patients due to less efficient 25(OH)D synthesis (2, 123).

1,25(OH)₂D

1,25(OH)₂D remains a milestone for the treatment of secondary HPT in CRF patients. It may be prescribed to patients with secondary HPT at every stage of CRF, although theoretically it should be used in the presence of normal serum levels of 25(OH)D. Exogenous administration of 1,25(OH)₂D becomes inevitable in patients with creatinine clearance below 15 mL/min (Table III), provided that the serum values of calcium, phosphate and PTH indicate its use (88). In CRF patients with creatinine clearance above 15 mL/min, 1,25(OH)₂D treatment may begun at the oral dose of 0.25 µg/day, according to the K/DOQI guidelines (88). Clinical trials aimed to study the outcome of 1,25(OH)₂D treatment showed that CRF patients, who had received 1,25(OH)₂D (0.25-0.5 µg/day per os) for one year, had lower circulating PTH and normal bone histology at the end of follow-up. A significantly higher bone turnover and lower bone mineral density were observed in patients who had not received 1,25(OH)₂D (95-97). Therefore, although protective against HPT, 1,25(OH)₂D treatment is likely to expose patients with early CRF to the risk of adynamic bone disease and may also accelerate renal function decline, probably due to vascular deposition of calcium-phosphate salts (98). However, the dosage of 0.125-0.25 µg/day was well tolerated in patients with mild-moderate CRF and appeared safe for bone and kid-

ney (99, 100).

1,25(OH)₂D is prescribed to hemodialysis patients as oral daily doses or intravenous (i.v.) oral thrice-weekly boli. The intermittent supplementation was studied for dialysis patients with the rationale to produce peaks of 1,25(OH)₂D concentration in blood. Pharmacokinetic considerations and experimental findings suggested that intermittent supplementation has the advantage of a greater PTH suppression and an easier patient compliance. In spite of this promising background, two trials comparing intermittent with daily administration of 1,25(OH)₂D in dialysis patients, showed no advantages of intermittent doses in terms of efficacy and safety (101, 102). About i.v. or oral boli, although K/DOQI guidelines conclude that i.v. boli were more efficient for the control of secondary HPT, a study was unable to find differences between the two administration routes (103). The expectation created by the introduction of the intermittent administration gave rise to a discussion about a possible "pharmacological parathyroidectomy", capable to abolish autonomous parathyroid secretion. Conversely, nephrologists learnt that excessive suppression of PTH was the main cause of adynamic bone disease (121, 122). Plasma concentrations of PTH between 150 and 300 pg/mL are now considered as the target for vitamin D treatment which has to be started only when PTH is above this range.

Vitamin D analogues

Several new vitamin D analogues have been developed and investigated with the rationale to treat secondary HPT decreasing the risk of hypercalcemia and hyperphosphatemia in CRF patients (83, 84). Vitamin D analogues have variable affinity for the components of the vitamin D system, including the vitamin D-binding protein and the nuclear VDR. Some of the effects are genomic and mediated through changes in the structural configuration of the vitamin D-VDR complex or in the affinity of the vitamin D-VDR complex for the key response elements in various target genes. There are currently three vitamin D analogues approved for use in CRF patients with secondary HPT in the US, 1,25-dihydroxy-22-oxavitamin D₃ (22-oxacalcitriol, OCT), 1,25-dihydroxy-19-norvitamin D₂ (19-norD₂), 1α-hydroxyvitamin D₂ (1αOHD₂). 19-norD₂ is approved for the treatment of secondary HPT in Italy.

OCT is a vitamin D₃ derivate, which differs from 1,25(OH)₂D₃ for the substitution of carbon 22 with an oxygen. Oral and i.v. preparations are available. Experience in rats showed that it is

Table III - Algorithm for treatment with 1,25(OH)₂D, OCT or 19-norD₂ in hemodialysis patients according to K/DOQI guidelines. Vitamin D should not be prescribed in patients with serum concentrations of calcium and phosphate higher than those indicated. The initial doses of vitamin D is related to plasma levels of PTH and then regulated on therapy outcome. K/DOQI guidelines consider the i.v. way as preferential with respect to oral way of administration.

To convert plasma calcium concentrations in mg/dL, multiply by 4; plasma phosphate concentrations in mg/dL, multiply by 3.1; calcium phosphate product in mg²/dL², multiply by 12.4.

Plasma PTH (pg/mL)	Plasma calcium (mmol/L)	Serum phosphate (mmol/L)	Calcium-phosphate product (mmol ² /L ²)	1,25(OH) ₂ D dose (µg after HD)	OCT dose (µg after HD)	19-norD ₂ dose (µg after HD)
300-600	<2.37	<1.78	<4.5	i.v.: 0.5-1 oral: 0.5-1	i.v.: 2.5-5	i.v.: 2 oral: 5
600-1000	<2.37	<1.78	<4.5	i.v.: 1-3 oral: 1-4	i.v.: 6-10	i.v.: 2-4 oral: 5-10
>1000	<2.5	<1.78	<4.5	i.v.: 3-5 oral: 3-7	i.v.: 10-15	i.v.: 4-8 oral: 10-20

less potent than 1,25(OH)₂D to suppress parathyroid glands and much less active on plasma calcium. OCT is commonly administered to hemodialysis (HD) patients three times a week (Table III). In the first trial, episodes of hypercalcemia occurred in 33% of patients and serum alkaline phosphatase decreased significantly after OCT therapy, suggesting the correction of high-turnover bone disease (104). The reduction in PTH and the increment of calcium and phosphate were dose-dependent. The initial OCT oral dose was 5 µg three times a week when plasma iPTH concentration was 300-500 pg/mL and 10 µg three times a week when iPTH was above 500 pg/mL (105). OCT was also successfully used in CRF patients with creatinine clearance above 15 mL/min at the oral dose of 1-5 µg/day with no deleterious effects on renal function (106).

The different activity of OCT and 1,25(OH)₂D has been attributed to the different pharmacokinetics of the two compounds. OCT affinity for VDR and vitamin D-binding protein is lower than that of 1,25(OH)₂D and, thus, it is rapidly cleared from the blood. OCT treatment is also associated with the inhibition of renal 1α-hydroxylase and a consequent reduction in serum levels of 1,25(OH)₂D. The short half-life and the 1,25(OH)₂D deficiency explain the scarce effect of OCT on intestine and bone and, ultimately, on calcium concentrations. On the other hand, OCT has a prolonged activity on parathyroid glands, where it is retained in the nuclei (107-108).

19-norD₂ has the side chain of vitamin D₂ and differs from 1,25(OH)₂D for the lack of carbon 22. It was the first analog approved for use in CRF patients and is available in Italy for i.v. administration, commonly three times a week after hemodialysis (Table III). It may be dosed according to a 4:1 ratio to 1,25(OH)₂D, even though a ratio 3:1 was proposed for a safer approach (109). Thus, in patients already taking 1,25(OH)₂D, the initial doses of 19-norD₂ can be calculated as four times that of 1,25(OH)₂D. Alternatively, its initial dosage may be calculated from circulating PTH concentrations (PTH concentration / 80) or body weight (0.04 µg/kg of dry body weight) (110). Efficacy of 19-norD₂ and 1,25(OH)₂D was compared in a randomized double-blind trial. Reduction in plasma PTH was more rapid in patient taking 19-norD₂, although the PTH reduction target (decrease of 50% of the baseline plasma concentration) was obtained in a similar proportion of patients after 20 weeks of treatment (79% with 19-norD₂ and 69% 1,25(OH)₂D). Severe hyperphosphatemia (above 2.58 mmol/L or 8 mg/dL) occurred in 9% of patients treated with 19-norD₂ and 14% of patients treated with 1,25(OH)₂D, while no differences in plasma calcium levels were observed (111). In another randomized double-blind study comparing 19-norD₂ with 1,25(OH)₂D, hypercalcemia and/or calcium-phosphate product above 6 mmol²/L² (75 mg²/dL²) occurred at least once in 68% of patients during treatment with 19-norD₂ and 64% with 1,25(OH)₂D (112). Conversely, persistent hypercalcemia was observed in 18% of patients treated with 19-norD₂ and 33% with 1,25(OH)₂D. These trials showed that 19-norD₂ was able to control PTH levels reducing the occurrence of hypercalcemia and hyperphosphatemia in comparison with 1,25(OH)₂D. Therefore, the control of PTH secretion is obtained with lower calcium-phosphate product with 19-norD₂ (111, 112). This reduces the risk of vascular calcification and may contribute to the increased survival observed in hemodialysis patients undergoing 19-norD₂ therapy compared with those taking 1,25(OH)₂D (113). Furthermore, in patients taking 19-norD₂ a reduced hospitalization rate was observed (114). However, the values of these studies about survival in patients treated with 19-norD₂ was limited by the fact they were retrospective and not randomized.

The limited hypercalcemic effect of 19-norD₂ results from a lower affinity for VDR in intestine and bone. In addition, 19-norD₂ causes a selective vitamin D deficiency and a reduction

in intestinal calcium absorption (115, 116).

1αOHD₂ is a prodrug activated by 25-hydroxylation in the liver. It may be administered daily or three times a week, orally or i.v. Clinical trials demonstrated a good efficacy of 1αOHD₂ in the treatment of secondary HPT in hemodialysis patients. 1αOHD₂, prescribed at the initial oral doses of 4 µg daily or three times a week, normalized PTH concentrations in 88% of patients, but hypercalcemia and hyperphosphatemia complicated the treatment in 42% and 75% of patients respectively (117). This high rate of hypercalcemia was not observed in other studies, although an increase in serum phosphate and calcium invariably occurred. In a larger trial including 99 hemodialysis patients, 1αOHD₂ was prescribed at the oral dose of 10 µg three times a week. Hypercalcemia was observed in 15% of patients and hyperphosphatemia in 19%. Plasma PTH decreased by 45% from baseline levels and became normal in 83% of patients (118). Intravenous and oral preparations of 1αOHD₂ are available. The increase of calcium and phosphate was lower with i.v. than oral administration, but the induced decline of PTH was similar. 1αOHD₂ was also used at the dose of 1-3 µg/day in non-uremic CRF patients. Its use was safe and effective in decreasing plasma PTH and increased bone mineral density (119).

The lower hypercalcemic effect of 1αOHD₂ with respect to 1αOHD₃ has been attributed to the production of 24-hydroxylated metabolites with less hypercalcemic activity (120).

Conclusions

The modern strategies to prevent secondary HPT in CRF patients give great relevance to vitamin D replacement therapy, which requires to take into account the stage of CRF, the underlying renal disorder, the levels of circulating PTH, the condition of the bone, the vitamin D stores, the parameters of bone turnover and the values of calcium and phosphate in serum.

The administration of 1,25(OH)₂D or its analogues is unavoidable in dialysis patients when secondary HPT is often already established. Its association with 25(OH)D or ergocalciferol or cholecalciferol may be beneficial for the bones, if a deficit of vitamin D stores is present (serum levels of 25(OH)D below 20-30 ng/mL). Therapeutic use of 25(OH)D or ergocalciferol is indicated in earlier stages of CRF to support renal 1α-hydroxylase activity. At these stages, low doses of 1,25(OH)₂D may also be employed to prevent HPT since they are not harmful for renal function. The dose of vitamin D metabolites should be titrated on serum concentrations of calcium and phosphate, to avoid an excessively high calcium-phosphate product, and on serum PTH concentrations, to avoid excessive PTH suppression and an adynamic condition of the bone.

The aim of vitamin D replacement therapy is to prevent HPT since the early stages of CRF, because parathyroid hyperplasia and osteodystrophy cannot be completely reverted once developed. The attention of nephrologists has largely focused on dialysis patients and, unfortunately, few studies have analyzed the outcome of vitamin D therapy in non-uremic patients. Because of the lack of clinical studies in this population no guidelines are available on when to start vitamin D replacement therapy in CRF patients. Therefore, it is likely that HPT and osteodystrophy are undertreated in a significant proportion of CRF patients. We have tried to summarize the criteria proposed in the current literature: one common criteria is that vitamin D therapy should be considered when serum 25(OH)D concentration is below 30 ng/mL. Instead, we do not know when 1,25(OH)₂D can be safely prescribed to non-uremic patients and whether there are additional benefits by its association with 25(OH)D or ergocalciferol.

On the other hand, vitamin D therapy is not without problems, the first of which being vascular calcification. Again, we should

make every effort to prevent vascular calcification since the early stages of CRF, because calcification is a non-reversible lesion. The K/DOQI guidelines provide some criteria to minimize the risk of calcium phosphate precipitation in terms of target serum concentration of calcium and phosphate. However, our methods to fight calcification are limited and even the calcium-phosphate product provides insufficient information.

The therapeutical strategies for secondary HPT are now changing. The availability of 1,25(OH)₂D analogues warrants inhibition of parathyroid glands with lower effect on calcium and phosphate levels, and perhaps reduces mortality of dialysis patients. In the near future 1,25(OH)₂D analogues will be combined with a new drug, cinacalcet, that directly inhibits PTH secretion and parathyroid proliferation. Potentially, the association of cinacalcet with 1,25(OH)₂D or its analogues is very promising because of the opposite effects of the two drugs on plasma calcium levels.

References

1. Reinhart TA, Ramberg CF, Horst RL. Comparison of receptor binding, biological activity, and in vivo tracer kinetics for 1,25-dihydroxyvitamin D₃ and 1,25-dihydroxyvitamin D₂, and its 24 epimer. *Arch Biochem Biophys*. 1989;273:64-71.
2. Cunningham J, Makin H. How important is vitamin deficiency in uremia? *Nephrol Dial Transplant*. 1997;12:16-18.
3. Khaw KT, Sneyed MJ, Compston J. Bone density, parathyroid hormone and 25-hydroxyvitamin D concentrations in middle aged women. *BMJ*. 1992;305:373-376.
4. McKenna MJ, Freaney R. Secondary hyperparathyroidism in the elderly: means to defining hypovitaminosis D. *Osteoporos Int*. 1998;8:S3-S6.
5. Cannata JB, Alonso CG. Vitamin D deficiency: a neglected aspect of disturbed calcium metabolism in renal failure. *Nephrol Dial Transplant*. 2002;17:1875-1878.
6. Ghazali A, Fardellone P, Pruna A, Atik A, Achard JM, Oprisiu R, Brazier M, Remond A, Moriniere P, Garabedian M, Eastwood J, Fournier A. Is low 25-(OH)vitamin D a major risk factor for hyperparathyroidism and Looser's zones independent of calcitriol. *Kidney Int*. 1999;55:2169-2177.
7. Wang M, Hercz Z, Sherrard DJ, Segre GV, Pei Y. Relationship between intact PTH 1-84 parathyroid hormone and bone histomorphometry parameters in dialysis patients without aluminum toxicity. *Am J Kidney Dis*. 1995;26:836-844.
8. Le Boff MS, Kholmeier L, Hurwitz S, Franklin J, Wright J, Glowacki J. Occult vitamin D deficiency in postmenopausal US women with acute hip fracture. *JAMA*. 1999;281:1505-1511.
9. Bischoff-Ferrari HA, Dietrich T, Orav EJ, Dawson-Hughes B. Positive Association between 25-Hydroxyvitamin D levels and bone mineral density: a population-based study of younger and older adults. *Am J Med*. 2004;116:634-639.
10. Collins D, Jasani C, Fogelman I, Swaminathan R. Vitamin D and bone mineral density. *Osteoporos Int*. 1998;8:110-114.
11. Thomas MK, Lloyd-Jones DM, Thadhani RI, Shaw AC, Deraska DJ, Kitch BT, Vamvakas EC, Dick IM, Prince RL, Finkelstein JS. Hypovitaminosis D in medical inpatients. *N Engl J Med*. 1998;338:777-783.
12. Coen G, Mantella D, Manni M, Balducci A, Nofroni I, Sardella D, Ballanti P, Bonucci E. 25-hydroxyvitamin D levels and bone histomorphometry in hemodialysis renal osteodystrophy. *Kidney Int*. 2005;68:1840-1845.
13. Lin R, White JH. The pleiotropic actions of vitamin D. *Bioessays*. 2004;26:21-28.
14. Zittermann A, Schleithoff SS, Tenderich G, Berthold HK, Korfer R, Stehle P. Low vitamin D status: a contributing factor in the pathogenesis of congestive heart failure? *J Am Coll Cardiol*. 2003;41:105-112.
15. Park CW, Oh YS, Shin YS, Kim CM, Kim YS, Kim SY, Choi EJ, Chang YS, Bang BK. Intravenous calcitriol regresses myocardial hypertrophy in hemodialysis patients with secondary hyperparathyroidism. *Am J Kidney Dis*. 1999;33:73-81.
16. Li YC, Kong J, Wei M, Chen ZF, Liu SQ, Cao LP. 1,25-dihydroxyvitamin D(3) is a negative endocrine regulator of the renin-angiotensin system. *J Clin Invest*. 2002;110:229-238.
17. Watson KE, Abrolat ML, Malone LL, Hoeg JM, Doherty T, Detrano R, Demer LL. Active serum vitamin D levels are inversely correlated with coronary calcification. *Circulation*. 1997;96:1755-1760.
18. Holick MF. Vitamin D: importance in the prevention of cancers, type 1 diabetes, heart disease, and osteoporosis. *Am J Clin Nutr*. 2004;79:362-371.
19. Holick MF. Evolution and function of vitamin D. *Recent Results Cancer Res*. 2003;164:3-28.
20. Parfitt AM. The hyperparathyroidism of chronic renal failure: A disorder of growth. *Kidney Int*. 1997;52:3-9.
21. Silver J, Bar Sela S, and Naveh-Many T. Regulation of parathyroid cell proliferation. *Curr Op Nephrol Hypertens*. 1997;6:321-326.
22. Cozzolino M, Brancaccio D, Gallieni M, Galassi A, Slatopolsky E, Dusso A. Pathogenesis of parathyroid hyperplasia in renal failure. *J Nephrol*. 2005;18:5-8.
23. Denda M, Finch J, Slatopolsky E. Phosphorus accelerated the development of parathyroid hyperplasia and secondary hyperparathyroidism in rats with renal failure. *Am J Kidney Dis*. 1996;28:596-602.
24. Tominaga Y, Tsuzuki T, Uchida K, Haba T, Otsuka S, Ichimori T, Yamada K, Numano M, Tanaka Y, Takagi H. Expression of Prad1/cyclin D1, retinoblastoma gene product, and Ki67 in parathyroid hyperplasia caused by chronic renal failure versus primary adenoma. *Kidney Int*. 1999;55:1375-1383.
25. Brown AJ, Zhong M, Ritter C, Brown EM, Slatopolsky E. Loss of calcium responsiveness in cultured bovine parathyroid cells is associated with decreased calcium sensing receptor expression. *Biochem Biophys Res Commun*. 1995;212:861-867.
26. Delmez JA, Slatopolsky E. Hyperphosphatemia: its consequences and treatment in patients with chronic renal disease. *Am J Kidney Dis*. 1992;19:303-317.
27. Rodriguez M, Almaden Y, Hernandez A, and Torres A. Effect of phosphate on the parathyroid gland: direct and indirect? *Curr Opin Nephrol Hypertens*. 1996;5:321-328.
28. Portale AA, Halloran BP, Morris RC Jr. Physiologic regulation of the serum concentration of 1,25-dihydroxyvitamin D by phosphorus in normal men. *J Clin Invest*. 1989;83:1494-1499.
29. Almaden Y, Canalejo A, Hernandez A, Ballesteros E, Garcia-Navarro S, Torres A, Rodriguez M. Direct effect of phosphorus on PTH secretion from whole rat parathyroid glands in vitro. *J Bone Miner Res*. 1996;11:970-976.
30. Nielsen PK, Feldt-Rasmussen U and Olgaard K. A direct effect in vitro of phosphate on PTH release from bovine parathyroid tissue slices but not from dispersed parathyroid cells. *Nephrol Dial Transplant*. 1996;11:1762-1768.
31. Moallem E, Kilav R, Silver J, and Naveh-Many T. RNA-Protein binding and post-transcriptional regulation of parathyroid hormone gene expression by calcium and phosphate. *J Biol Chem*. 1998;273:5253-5259.
32. Wang Q, Palnitkar S, Parfitt AM. Parathyroid cell proliferation in normal human parathyroid tissue: implications for the pathogenesis of hyperparathyroidism. *Clin Endocrinol*. 1997;46:343-349.
33. Dusso AS, Pavlopoulos T, Naumovich L, Lu Y, Finch J, Brown AJ, Morrissey J, Slatopolsky E. p21waf1 and TGF α mediate dietary phosphate-regulation of parathyroid cell growth. *Kidney Int*. 2001;59:855-865.
34. Kumar V, Bustiin SA, McKay IA. Transforming growth factor alpha. *Cell Biol Int*. 1995;19:373-388.
35. Gogusev J, Duchambon P, Stoeremann-Chopard C, Giovannini M, Sarfati E, Druke TB. De novo expression of transforming growth factor-alpha in parathyroid gland tissue of patients with primary or secondary uraemic hyperparathyroidism. *Nephrol Dial Transplant*. 1996;11:2155-2162.
36. Cozzolino M, Lu Y, Finch J, Slatopolsky E, Dusso AS. p21WAF1

- and TGF α mediate parathyroid growth arrest by vitamin D and high calcium. *Kidney Int.* 2001;60:2109-2117.
37. Wong ST, Winchell LF, McCune BK, Earp HS, Teixido J, Masague J, Herman B, Lee DC. The TGF- α precursor expressed on the cell surface binds to the EGF receptor on adjacent cells, leading to signal transduction. *Cell.* 1989;56:495-506.
 38. Sadler GP, Morgan JM, Jasani B, Douglas-Jones A, Wheeler MH. Epidermal growth factor receptor status in hyperparathyroidism: immunocytochemical and in situ hybridization study. *World J Surg.* 1996;20:736-743.
 39. Dusso A, Cozzolino M, Lu Y, Sato T, Slatopolsky E. 1,25-Dihydroxyvitamin D downregulation of TGF α /EGFR expression and growth signaling: a mechanism for the antiproliferative actions of the sterol in parathyroid hyperplasia of renal failure. *J Steroid Biochem Mol Biol.* 2004;89-90:507-511.
 40. Cozzolino M, Lu Y, Sato T, Yang J, Suarez IG, Brancaccio D, Slatopolsky E, Dusso AS. A critical role for enhanced-TGF α and EGFR expression in the initiation of parathyroid hyperplasia in experimental renal disease. *Am J Physiol Renal Physiol.* 2005;Jul 5;[Epub ahead of print].
 41. Brown EM. Calcium receptor and regulation of parathyroid hormone secretion. *Rev Endocr Metab Disord.* 2000;1:307-315.
 42. Ho C, Conner DA, Pollak MR, Ladd DJ, Kifor O, Warren HB, Brown EM, Seidman JG, Seidman CE. A mouse model of human familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. *Nat Genet.* 1995;11:389-394.
 43. Li YC, Amling M, Pirro AE, Priemel M, Meuse J, Baron R, Delling G, Demay MB. Normalization of mineral on homeostasis by dietary means prevents hyperparathyroidism, rickets, and osteomalacia, but not alopecia in vitamin D receptor-ablated mice. *Endocrinology.* 1998;139:4391-4396.
 44. Akizawa T, Fukagawa M. Modulation of parathyroid cell function by calcium ion in health and uremia. *Am J Med Sci.* 1999;317:358-362.
 45. Nemeth EF. Calcimimetic and calcilytic drugs: just for parathyroid cells? *Cell Calcium.* 2004;35:283-289.
 46. Naveh-Many T, Rahaminov R, Livni N, Silver J. Parathyroid cell proliferation in normal and chronic renal failure rats. The effect of calcium, phosphate, and vitamin D. *J Clin Invest.* 1995;96:1786-1793.
 47. Garfia B, Canadillas S, Canalejo A, Luque F, Siendones E, Quesada M, Almaden Y, Aguilera-Tejero E, Rodriguez M. Regulation of parathyroid vitamin D receptor expression by extracellular calcium. *J Am Soc Nephrol.* 2002;13:2945-2952.
 48. Brown AJ, Ritter CS, Finch JL, Slatopolsky E. Decreased calcium-sensing receptor expression in hyperplastic parathyroid glands of uremic rats: role of dietary phosphate. *Kidney Int.* 1999;55:1284-1292.
 49. Yano S, Sugimoto T, Tsukamoto T, Chihara K, Kobayashi A, Kitazawa S, Maeda S, Kitazawa R. Association of decreased calcium-sensing receptor expression with proliferation of parathyroid cells in secondary hyperparathyroidism. *Kidney Int.* 2000;58:1980-1986.
 50. Amling M, Priemel M, Holzmann T, Chapin K, Rueger JM, Baron R, Demay MB. Rescue of the skeletal phenotype of vitamin D receptor-ablated mice in the setting of normal mineral ion homeostasis: formal histomorphometric and biochemical analyses. *Endocrinology.* 1999;140:4982-4987.
 51. Motellon JL, Jimenez FJ, de Miguel F, Jaras MJ, Diaz A, Hurtado J, Esbrit P. Relationship of plasma bone cytokines with hypercalcemia in cancer patients. *Clin Chim Acta.* 2000;302:59-68.
 52. Kremer R, Bolivar I, Goltzman D, Hendly GN. Influence of calcium and 1,25-dihydroxycholecalciferol on proliferation and proto-oncogene expression in primary cultures of bovine parathyroid cells. *Endocrinology.* 1989;125:935-941.
 53. Szabo A, Merke J, Beier E, Mall G, Ritz E. 1,25(OH) $_2$ vitamin D $_3$ inhibits parathyroid cell proliferation in experimental uremia. *Kidney Int.* 1989;35:1049-1056.
 54. Naveh-Many T and Silver J. Regulation of parathyroid hormone gene expression by hypocalcemia, hypercalcemia, and vitamin D in the rat. *J Clin Invest.* 1990;86:1313-1319.
 55. Tokumoto M, Tsuruya K, Fukuda K, Kanai H, Kuroki S, Hirakata H. Reduced p21, p27 and vitamin D receptor in the nodular hyperplasia in patients with advanced secondary hyperparathyroidism. *Kidney Int.* 2002;62:1196-1207.
 56. Cordero JB, Cozzolino M, Lu Y, Vidal M, Slatopolsky E, Stahl PD, Barbieri MA, Dusso A. 1,25-dihydroxyvitamin D downregulates cell membrane growth- and nuclear growth-promoting signals by the epidermal growth factor receptor. *J Biol Chem.* 2002;277: 38965-38971.
 57. Martinez I, Saracho R, Montenegro J, Llach F. A deficit of calcitriol synthesis may not be the initial factor in the pathogenesis of secondary hyperparathyroidism. *Nephrol Dial Transplant.* 1996;11 Suppl 3:22-28.
 58. Reichel H, Deibert B, Schmidt-Gayk H, Ritz E. Calcium metabolism in early chronic renal failure, implications for the pathogenesis of hyperparathyroidism. *Nephrol Dial Transplant.* 1991;6:162-169.
 59. Eastwood JB, Harris E, StampTCB, De Wardener HE. Vitamin D deficiency in the osteomalacia of chronic renal failure. *Lancet.* 1976;2:1209-1221.
 60. Van der Wielen RP, Lowik MR, Van den Berg H, deGroot L, Haller J, Mereiras W, Van Staveren WA. Serum vitamin D concentrations among elderly people in Europe. *Lancet.* 1995;346:207-210.
 61. Scharla SH. Prevalence of subclinical vitamin D deficiency in different European countries. *Osteoporos Int.* 1998;8:s7-s12.
 62. Ishimura E, Nishizawa Y, Inaba M, Matsumoto N, Emoto M, Kawagishi T, Shoji S, Okuno S, Kim M, Miki T, Morii H. Serum levels of 1,25-dihydroxyvitamin D, 24,25-dihydroxyvitamin D, and 25-hydroxyvitamin D in non dialyzed patients with chronic renal failure. *Kidney Int.* 1999;55:1019-1027.
 63. Messa P, Vallone C, Meoni G, Geatti O, Turrin D, Passoni N, Cruciani A. Direct in vivo assessment of parathyroid hormone calcium relationship curve in renal patients. *Kidney Int.* 1994;46:1713-1720.
 64. Slovik DM, Adams JS, Neer RS. Deficient production of 1,25-dihydroxyvitamin D in elderly osteoporotic patients. *N Engl J Med.* 1981;305:372-374.
 65. Papapoulos SE, Clemens TL, Fraher LJ, Gleed J, O'Riordan JL. Metabolites of vitamin D in human vitamin D deficiency. Effect of vitamin D or 1,25-dihydroxycholecalciferol. *Lancet.* 1980;2:612-615.
 66. Battaille P, Achard JM, Fournier A, Boudailliez B, Westeel JD, El Espe N, Bergot C, Jans I, Lalau JD, Petit J, Henon G, Laval Jean-tet MA, Bouillon R, Sebert JL. Diet, vitamin D and vertebral mineral density in hypercalciuric calcium stone formers. *Kidney Int.* 1991;39:1193-1205.
 67. Cogan MG, Covey CM, Arief AI, Wisniewski A, Clark OH, Lazarowitz V, Leach W. Central nervous system manifestations of hyperparathyroidism. *Am J Med.* 1978;963-970.
 68. Mak RH, Bettinelli A, Turner C, Haycock GB, Chantler C. The influence of hyperparathyroidism on glucose metabolism in uremia. *J Clin Endocrinol Metab.* 1985;60:229-233.
 69. Elder G. Pathophysiology and recent advances in the management of renal osteodystrophy. *J Bone Miner Res.* 2002;17:2094-2105.
 70. Ganesh SK, Stack AG, Levin NW, Hulbert-Shearon T, Port FK. Association of elevated serum PO(4), Ca-PO(4) product, and parathyroid hormone with cardiac mortality risk in chronic hemodialysis patients. *J Am Soc Nephrol.* 2001;12:2131-2138.
 71. Cozzolino M, Dusso A, Slatopolsky E. Role of calcium-phosphate product and bone associated proteins on vascular calcification in renal failure. *J Am Soc Nephrol.* 2001;12:2511-2516.
 72. London GM. Cardiovascular calcifications in uremic patients: clinical impact on cardiovascular function. *J Am Soc Nephrol.* 2003; 14:S305-309.
 73. Blacher J, Safar ME, Guerin AP, Pannier B, Marchais SJ, London GM. Aortic pulse wave velocity index and mortality in end-stage renal disease. *Kidney Int.* 2003;63:1852-1860.
 74. Cozzolino M, Brancaccio D, Gallieni M, Slatopolsky E. Pathogenesis of vascular calcification in chronic kidney disease. *Kidney Int.* 2005;68(2):429-436.

75. Jakoby MG, Semenkovich CF. The role of osteoprogenitors in vascular calcification. *Curr Opin Nephrol Hypertens.* 2000;9:11-15.
76. Brancaccio D, Cozzolino M. The mechanism of calcium deposition in soft tissues. *Contrib Nephrol.* 2005;149:279-286.
77. Block GA, Port FK. Re-evaluation of risks associated with hyperphosphatemia and hyperparathyroidism in dialysis patients: recommendations for a change in management. *Am J Kidney Dis.* 2000;35:1226-1237.
78. London GM, Guerin AP, Marchais SJ, Metivier F, Pannier B, Adda H. Arterial media calcification in end-stage renal disease: impact on all-cause and cardiovascular mortality. *Nephrol Dial Transplant.* 2003;18:1731-1740.
79. Goodman WG, Goldin J, Kuizon BD, Yoon C, Gales B, Sider D, Wang Y, Chung J, Emerick A, Greaser L, Elashoff RM, Salusky IB. Coronary-artery calcification in young adults with end-stage renal disease who are undergoing dialysis. *NEJM.* 2000;342:1478-1483.
80. Cozzolino M, Dusso AS, Liapis H, Finch J, Lu Y, Burke SK, Slatopolsky E. The effects of sevelamer hydrochloride and calcium carbonate on kidney calcification in uremic rats. *J Am Soc Nephrol.* 2002;13:2299-2308.
81. Cozzolino M, Staniforth ME, Liapis H, Finch J, Burke SK, Dusso AS, Slatopolsky E. Sevelamer hydrochloride attenuates kidney and cardiovascular calcifications in long-term experimental uremia. *Kidney Int.* 2003;64:1653-1661.
82. Chertow GM, Burke SK, Raggi P. Treat to Goal Working Group. Sevelamer attenuates the progression of coronary and aortic calcification in hemodialysis patients. *Kidney Int.* 2002;62:245-252.
83. Okazaki T, Zajac JD, Igarashi T, Ogata E, Kroenenberg HM. Negative regulatory elements in the human parathyroid hormone gene. *J Biol Chem.* 1991;266:21903-21910.
84. Kifor O, MacLeod RJ, Diaz R, Bai M, Yamaguchi T, Yao T, Kifor I, Brown EM. Regulation of MAPkinase by calcium-sensing receptor in bovine parathyroid and CaR-transfected HEK293 cells. *Am J Physiol Renal Physiol.* 2001;280:291-302.
85. Canaff L, Hendy G. Human calcium-sensing receptor gene. Vitamin D response elements in promoter P1 and P2 confer transcriptional responsiveness to 1,25-dihydroxyvitamin D. *J Biol Chem.* 2002;277:30337-30350.
86. Yalcindag C, Silver J, Naveh-Mary. Mechanism of increased parathyroid hormone mRNA in experimental uremia: roles of protein RNA binding and RNA degradation. *J Am Soc Nephrol.* 1999;10:2562-2568.
87. Okazaki T, Ando K, Igarashi T, Ogata E, Fujita T. Conserved mechanism of negative gene regulation by extracellular calcium parathyroid hormone gene versus atrial natriuretic polypeptide gene. *J Clin Invest.* 1992;89:1268-1273.
88. K/DOQI Clinical Practice Guidelines for Bone Metabolism and Disease in Chronic Kidney Disease. *Am J Kidney Dis.* 2003;42 (suppl. 3):63-200.
89. Massry SG, Coburn JW, Lee DB, Jowsey J, Kleeman CR. Skeletal resistance to parathyroid hormone in renal failure. Studies in 105 human subjects. *Ann Intern Med.* 1973;78:357-364.
90. Disthabanchong S, Hassan H, McConkey CL, Martin KJ, Gonzalez EA. Regulation of PTH1 receptor expression by uremic ultrafiltrate in UMR 106-101 osteoblast-like cells. *Kidney Int.* 2004;65:897-903.
91. Hercz G, Pei Y, Greenwood C, Manuel A, Saiphoo C, Goodman WG, Segre GV, Fenton S, Sherrard DJ. Aplastic osteodystrophy without aluminum: the role of "suppressed" parathyroid function. *Kidney Int.* 1993;44:860-866.
92. Coco M, Rush H. Increased incidence of hip fractures in dialysis patients with low serum parathyroid hormone. *Am J Kidney Dis.* 2000;36:1115-1121.
93. Cournot-Witmer G, Zingraff J, Plachot JJ, Escaig F, Lefevre R, Boumati P, Bourdeau A, Garabedian M, Galle P, Bourdon R, Druke T, Balsan S. Aluminum localization in bone from hemodialyzed patients: relationship to matrix mineralization. *Kidney Int.* 1981;20:375-378.
94. Birge SJ, Haddad JG. 25-hydroxycholecalciferol stimulation of muscle metabolism. *J Clin Invest.* 1975;56:1100-1107.
95. Nordal KP, Dahl E. Low dose calcitriol versus placebo in patients with predialysis chronic renal failure. *J Clin Endocrinol Metab.* 1988;67:929-936.
96. Baker LR, Abrams L, Roe CJ, Faugere MC, Fanti P, Subayti Y, Malluche HH. 1,25(OH)₂D₃ administration in moderate renal failure. A prospective double-blind trial. *Kidney Int.* 1989;35:661-669.
97. Bianchi ML, Colantonio G, Campanini F, Rossi R, Valenti G, Ortolani S, Bucciatti G. Calcitriol and calcium carbonate therapy in early chronic renal failure. *Nephrol Dial Transplant.* 1994;9:1595-1599.
98. Christiansen C, Rodbro P, Christiansen MS, Hartnack B. Is 1,25-dihydroxycholecalciferol harmful to renal function in patients with chronic renal failure? *Clin Endocrinol.* 1981;15:229-236.
99. Ritz E, Kuster S, Schmidt-Gayk H, Stein G, Scholz C, Kraatz G, Heidland A. Low doses calcitriol prevents the rise in 1,84-iPTH without affecting serum calcium and phosphate in patients with moderate renal failure. *Nephrol Dial Transplant.* 1995;10:2228-2234.
100. Baker LR, Abram L, Roe CJ, Faugere MC, Fanti P, Subayti Y, Malluche HH. 1,25(OH)₂D₃ administration in moderate renal failure: a prospective double-blind trial. *Kidney Int.* 1989;35:661-669.
101. Herrmann P, Ritz E, Schmidt-Gayk H, Schafer I, Geyer J, Nonnast-Daniel B, Koch KM, Weber U, Horl W, Haas-Worle A. Comparison of intermittent and continuous oral administration of calcitriol in dialysis patients: a randomized prospective trial. *Nephron.* 1994;67:48-53.
102. Indridason OS, Quarles LD. Comparison of treatment for mild secondary hyperparathyroidism in hemodialysis patients. *Kidney Int.* 2000;57:282-292.
103. Levine BS, Song M. Pharmacokinetics and efficacy of pulse oral versus intravenous calcitriol in hemodialysis patients. *J Am Soc Nephrol.* 1996;7:488-496.
104. Akizawa T, Kurokawa K. Long-term clinical effect of maxacalcitol on hemodialysis patients with secondary hyperparathyroidism. *Clin Calcium.* 2002;12:781-788.
105. Akizawa T, Ohashi Y, Akiba T, Suzuki M, Nishizawa Y, Ogata E, Slatopolsky E, Kurokawa K. Dose-response study of 22-oxacalcitriol in patients with secondary hyperparathyroidism. *Ther Apher Dial.* 2004;8:480-491.
106. Coburn JW, Maung HM, Elangovan L, Germain J, Lindberg JS, Sprague SM, Williams ME, Bishop CW. Doxercalciferol safely suppress PTH levels in patients with secondary hyperparathyroidism associated with chronic kidney disease stages 3 and 4. *Am J Kidney Dis.* 2004;43:877-890.
107. Dusso AS, Negrea L, Finch J et al. The effect of 22-oxacalcitriol on serum calcitriol. *Endocrinology.* 1992;130:3129-3134.
108. Brown AJ, Finch JL, Lopez-Hilker S, Dusso A, Ritter C, Pernaletto N, Slatopolsky E. New active analogues of vitamin D with low calcemic activity. *Kidney Int.* 1990;29(suppl 29):22-27.
109. Lach F, Yudd M. Paricalcitol in dialysis patients with calcitriol resistant secondary hyperparathyroidism. *Am J Kidney Dis.* 2001;38(s5):45-50.
110. Martin KJ, Gonzales E, Lindberg J, Taccetta C, Amdahl M, Malhotra K, Llach F. Paricalcitol dosing according to body weight or severity of hyperparathyroidism: a vitamin D analogues for secondary hyperparathyroidism. A double-blind multicenter randomized trial. *Am J Kidney Dis.* 2001;38:s57-s63.
111. Sprague SM, Lerma E, McCormick D, Abraham M, Battle D. Suppression of parathyroid hormone secretion in hemodialysis patients: comparison of paricalcitol with calcitriol. *Am J Kidney Dis.* 2001;38(5 Suppl 5):S51-56.
112. Sprague SM, Llach F, Amdahl M, Taccetta C, Battle D. Paricalcitol versus calcitriol in the treatment of secondary hyperparathyroidism. *Kidney Int.* 2003;63:1483-1490.
113. Teng M, Wolf M, Lowrie E, Ofsthun N, Lazarus JN, Thadhani R. Survival of patients undergoing hemodialysis with paricalcitol or calcitriol therapy. *N Engl J Med.* 2003;349:446-456.
114. Dobrez DG, Mathes A, Amdahl M, Marx SE, Melnick JZ, Sprague SM. Paricalcitol-treated patients experience improved hospitaliza-

- tion outcomes compared with calcitriol-treated patients. *Nephrol Dial Transplant*. 2004;19:1174-1181.
115. Brown AJ, Finch J, Slatopolsky E. Differential effects of 19-nor-1,25-dihydroxyvitamin D(2) and 1,25-dihydroxyvitamin D(3) on intestinal calcium and phosphate transport. *J Lab Clin Med*. 2002; 139:279-284.
 116. Finch J, Brown AJ, Slatopolsky E. Differential effects of 1,25-dihydroxy-vitamin D3 and 19-nor-1,25-dihydroxy-vitamin D2 on calcium and phosphorus resorption in bone. *J Am Soc Nephrol*. 1999; 10:980-985.
 117. Tan AU Jr, Levine BS, Mazess RB, Kylo DM, Bishop CW, Knutson JC, Kleinman KS, Coburn JW. Effective suppression of parathyroid hormone by 1 α -hydroxy-vitamin D2 in hemodialysis patients with moderate to severe secondary hyperparathyroidism. *Kidney Int*. 1997;51:317-323.
 118. Franzao JM, Chesney RW, Coburn JW. Intermittent oral 1 α -hydroxyvitamin D2 is effective and safe for the suppression of secondary hyperparathyroidism in hemodialysis patients. *Nephrol Dial Transplant*. 1998;3:68-72.
 119. Rix M, Eskildsen O, Olgaard K. Effect of 18 months of treatment with alphacalcidol on bone in patients with mild to moderate chronic renal failure. *Nephrol Dial Transplant*. 2004;19:870-876.
 120. Mawer EB, Jones G, Davies M, Still PE, Byford V, Schroeder NJ, Makin HL, Bishop CW, Knutson JC. Unique 24-hydroxylated metabolites represent a significant pathway of metabolism of vitamin D2 in humans: 24-hydroxyvitamin D2 and 1,24-dihydroxyvitamin D2 detectable in human serum. *J Clin Endocrinol Metab*. 1998;83:2156-2166.
 121. Goodman WG, Ramirez JA, Belin TR, Chon Y, Gales B, Segre GV, Salusky IB. Development of adynamic bone in patients with secondary hyperparathyroidism after intermittent calcitriol therapy. *Kidney Int*. 1994;46:1160-1166.
 122. Schomig M, Ritz E. Management of disturbed calcium metabolism in uremic patients: 1. Use of vitamin D metabolites. *Nephrol Dial Transplant*. 2000;15(suppl 5):18-24.