

The genetics of response to estrogen treatment

Bente L. Langdahl

Department of Endocrinology and Metabolism Aarhus
University Hospital, Aarhus, Denmark

Address for correspondence:
Bente L. Langdahl MD, PhD, DMSc
Department of Endocrinology and Metabolism
Aarhus University Hospital
Tage-Hansens gade 2
DK-8000 Aarhus C
Denmark
Ph. +45 8949 7678
Fax +45 8949 7659
E-mail: bentlang@rm.dk

Summary

It has been demonstrated that the response to estrogen treatment in postmenopausal women shows considerable variability. It has been speculated that this at least partly could be determined by heritable factors.

The most obvious genes to investigate in this context are the estrogen receptor genes. It has been demonstrated that women with short alleles of the TA-repeat polymorphism in the estrogen receptor α gene respond to hormone treatment with greater increases in bone mass at the lumbar spine. Also the two polymorphisms in the first intron of the same gene have been found to be associated with the response to estrogen. Several studies have found that women carrying the P- and the X-alleles respond to hormone therapy with greater increases in bone mass and sustain fewer fractures.

Polymorphisms in the collagen type I α 1 have been found to influence BMD. Conflicting results have been obtained with respect to the influence of these genetic variants on postmenopausal bone loss and response to hormone treatment.

Furthermore, two polymorphisms in the promoter of the transforming growth factor β gene and one polymorphism in the first exon of the osteoprotegerin gene have been demonstrated to interact with the response to hormone treatment in early postmenopausal women.

The above mentioned results are obtained from relatively small studies and needs confirmation before the information can be used in the clinic.

KEY WORDS: estrogen, genetics, bone mass, osteoporosis.

Introduction

The differences in fracture risk between men and women are predominantly caused by the difference in bone size, men having bigger bones than women (1) and the abrupt withdrawal of estrogen at menopause in women. Estrogen deficiency, as seen in postmenopausal women, is associated with increased rate of bone loss and increased risk of osteoporotic fractures. Treatment with estrogen has therefore been used both as a

prophylactic measure to prevent development of osteoporosis and as treatment of existing osteoporosis. Several studies have shown that estrogen treatment in postmenopausal women has profound effects both on bone mineral density (BMD) and fracture risk (2, 3). These studies have also shown that there is a considerable variability between women in the response to estrogen treatment. Since several other menopause and estrogen related phenotypes, including postmenopausal bone loss, have been demonstrated to be influenced by genetic factors (4-11), it has also been speculated that the response to hormone replacement therapy (HRT) could be partly determined by heritable factors.

From a scientific point of view investigating this further would be of interest, since knowledge about the potential influence of genetic variants on response to hormone replacement treatment could lead to a deeper understanding of the importance of the genes and the functional importance of the genetic variants. From the patients point of view knowledge about the influence of genetic variants would be desirable. If genetic variants interfere with proper response to treatment, the patient might want a different treatment or no treatment. Furthermore, if genetic variants also increase the risk of side effects, the balance between positive and negative effects might change significantly and the patient may choose not to be treated. In the following some examples of interaction between response to HRT and genetic variants will be mentioned.

Hormones and their receptors

Estrogen receptors (ER) and aromatases

In the promoter of the *ER- α* gene a TA repeat polymorphism has been demonstrated (rs3138774) (12). The polymorphism is located 1174 base pairs upstream from the first exon. Short TA repeat alleles have been found to be associated with reduced BMD and a BMD-independent predictor of osteoporotic fractures (13, 14). In a study comprising 284 postmenopausal Korean women treated with equine estrogen alone or in combination with medroxyprogesterone for one year, Yim et al. found that women with short TA repeat alleles responded with greater increases in BMD at the lumbar spine but not at the femoral neck (15). Furthermore, women that lost bone mineral density, despite being compliant with the hormone replacement therapy, had significantly longer TA repeat alleles.

In the first intron 2 polymorphisms have been demonstrated, T to C 397 base pairs upstream from exon 2 identified by RFLP assay using *PvuII* (rs2234693) (16) and A to G 351 base pairs upstream from exon 2, identified by RFLP using *XbaI* (rs9340799) (17). These two polymorphisms are in strong linkage disequilibrium with each other, but also with the TA repeat polymorphism in the promoter. These polymorphisms have been examined separately and in combination and have in some studies been demonstrated to be associated with bone mass in men and women (14, 18, 19). However, the largest metaanalysis to date comprising 18,917 women and men, found no effect of the polymorphisms on BMD, but women car-

rying the XX genotype had reduced risk of fractures in general and vertebral fractures in particular (20).

Several studies have found associations between these polymorphisms and postmenopausal bone loss, however, not always with the same genotype (19, 21-25). The possible influence of the *PvuII* and *XbaI* polymorphisms on changes in bone mass during hormone replacement therapy in postmenopausal women has been examined in several studies. In an American study comprising 79 women treated with hormone replacement therapy for 3 years, Rapuri et al. found that women with the PP and the XX genotypes responded with higher increases in BMD at the lumbar spine, the femoral neck and total body, however, the increase was only significantly different at the total body for women with the XX genotype (24). In a study comprising 124 Thai women, it was demonstrated that after one year of treatment with low dose equine estrogen, women carrying the P allele responded with greater increase in BMD at the lumbar spine, compared with women with the PP genotype. No genotype dependent differences were found in women treated with higher doses or at the hip for any dose (26). Several other investigators found no association between these genotypes and bone mass response to hormone replacement therapy (21, 22, 25, 27-31). In a Finnish study, comprising 151 early postmenopausal women who were treated with estradiol for 5 years, Salmen et al. found that women carrying the P-allele sustained significantly fewer fractures compared with women with the pp genotype (32). Weel et al. have found that *PvuII* alleles are associated with early natural menopause and the risk of hysterectomy due to metrorrhagia and uterus myomatosis (33).

A tetranucleotide repeat polymorphism has been demonstrated in intron 4 of the aromatase CYP-19 gene. The effect of this polymorphism on BMD has been examined in several studies. Some studies found that few repeats were associated with reduced bone mass and increased risk of osteoporotic fractures (34, 35). In a prospective study, the Danish Osteoporosis Prevention Study (DOPS) comprising approximately 1800 early postmenopausal women of whom 440 received continuous treatment with estradiol for 5 years, Tofteng et al. found that response to hormone replacement therapy revealed an allele dose effect of the long allele on BMD at the lumbar spine, the hip and the forearm (36).

Vitamin D receptor (VDR)

The most studied genetic variants in the field of bone mass and osteoporotic fractures are located in the 3' end of the VDR. They comprise two variants in intron 8: G to A (*BsmI*) (rs1544410) and G to T (*ApaI*) (rs17879735), T¹⁰⁵⁵-C in exon 9 (*TaqI*) (rs17880009) and a poly-A microsatellite along with many other SNPs further down the 3'UTR of the gene (37-40). The b, a and T alleles are in strong linkage disequilibrium forming a baT haplotype. This baT haplotype is also strongly coupled to the long polyA repeat sequence. Two meta-analyses from 1996 and 1999 found that the 3' polymorphisms were weakly associated with bone mass, but non-genetic factors and genetic heterogeneity may interfere with the detection of the effects (41, 42). The largest meta-analysis comprising 26,242 women and men was performed using individual data and found no association between the C²T or the 3' polymorphisms and BMD or fracture risk, however, an association between BMD and the *Cdx2* polymorphism (rs17883966) was found (43).

Kurabayashi et al. followed 82 postmenopausal women treated with equine estrogen for at least 3 years. Initially women with the TT genotype responded with significantly greater increases in lumbar spine BMD (27), however, after 3 years no difference

between the two groups of women could be demonstrated (30). In a small American study comprising 54 women treated with low-dose equine estrogen for 3.5 years, Deng et al. found that women with the BB genotype responded better (22). Giguere et al. found no effect of the *BsmI* polymorphism on changes in QUS (quantitative ultrasound) during hormone replacement therapy (29). However, a subgroup of women with the bb genotype and the PP genotype of the *PvuII* polymorphisms in the *ER-α* gene had a more pronounced response to long-term hormone replacement therapy than women with the other possible combinations of these two genotypes.

Collagen, matrix proteins and other genes that influence bone structure

Collagen type Iα1 and methyltetrahydrofolate reductase (MTHFR)

The variant T-allele of the Sp1 polymorphism has been found to be associated with reduced BMD and increased fracture risk (44-46). Meta-analyses of which the largest comprised 20,786 women and men have confirmed the association of the T-allele with reduced bone mass and increased risk of osteoporotic fractures (47-49). In a Scottish study comprising 239 early postmenopausal women treated with estradiol for 5-7 years, no difference in the effect of the treatment on BMD was found between women with different Sp1 genotypes (50). In a small Turkish study comprising 111 early postmenopausal women, women carrying the normal genotype responded better to 18 months of hormone replacement therapy (51). In the Danish Osteoporosis Prevention Study, the promoter polymorphism G⁻¹⁹⁹⁷-T (rs1107946) was associated with reduced bone mass, but none of the investigated polymorphisms affected the response to HRT (52).

A point mutation in the MTHFR gene, causing an alanine to valine substitution, the C⁶⁷⁷T mutation (rs1801133), gives rise to a thermolabile variant of MTHFR with reduced activity (53), resulting in elevated levels of circulating homocysteine. Elevated plasma levels of homocysteine interfere with the cross-linking of collagen, a process that is essential in the formation of the triplehelix-structure of collagen type I during bone formation (54). The effect of this polymorphism on BMD has been examined in several studies and most studies have found that the TT genotype is associated with reduced BMD (55, 56) and increased risk of osteoporotic fractures (56, 57). However, in a study comprising 473 early postmenopausal Danish women, the BMD response to 5 years of hormone replacement therapy was not influenced by this polymorphism (56).

Other genes

Transforming Growth Factor-β (TGF-β), osteoprotegerin (OPG), LDL receptor-related peptide (LRP)-5 and -6 and Sclerostin (SOST)

Several polymorphisms have been identified in the *TGF-β1* gene (58-61). Some of these polymorphisms have been found to be associated with bone mass or fracture risk: C⁻¹³⁴⁸-T (60, 62), T²⁹-C (63, 64) and T⁸⁶¹⁻²⁰-C (60, 61). In a large-scale metaanalysis no effect of these polymorphisms on either BMD or risk of osteoporotic fractures could be demonstrated. Furthermore, no interaction between these polymorphisms and postmenopausal hormone replacement therapy was found (65). However, in the Danish Osteoporosis Prevention Study,

the C⁻¹³⁴⁸-T and T²⁹-C polymorphisms were associated with early postmenopausal bone loss and response to hormone replacement therapy (66).

Osteoprotegerin (OPG) is a decoy receptor for RANKL and a competitive inhibitor of osteoclast recruitment and activity (67). Several polymorphisms have been demonstrated in the *OPG* gene, some of these have been associated with changes in bone mass and fracture risk (68-71). In the Danish Osteoporosis Prevention Study, we found that two of the promoter polymorphisms; A¹⁶³-G and T²⁴⁵-G were associated with perimenopausal bone mass, but not with postmenopausal bone loss or response to hormone replacement therapy. Another polymorphism: G¹¹⁸¹-C located in the first exon interacted significantly with the response to hormone replacement therapy over 5 and 10 years (72).

Several disease associated mutations (73, 74), but also polymorphisms have been identified in the LDL receptor-related peptide (LRP)-5 and -6. Two polymorphisms in the *LRP-5* gene: Val⁶⁶⁷-Met and Ala¹³³⁰-Val have consistently been associated with reduced bone mass and increased fracture risk (75). The effect on BMD was confirmed in the Danish Osteoporosis Prevention Study. However, no effect of the polymorphisms on early postmenopausal bone loss or response to hormone replacement therapy could be demonstrated (76).

The *SOST* gene encodes sclerostin that is produced by the osteocytes and is a strong inhibitor of bone formation. Loss-of-function mutations cause sclerosteosis (77) and van Buchem's disease (78). Common polymorphisms in the *SOST* gene are associated with changes in bone mass (79). These effects on bone mass were confirmed in perimenopausal Danish women. Furthermore, the SRP9 polymorphism was also associated with early postmenopausal bone loss (80).

Discussion and future aspects

The interests in unraveling the genetics behind the variable response to hormone replacement therapy with respect to changes in bone mass, reduction in fracture risk and increase in risk of breast cancer and cardiovascular disease are several. For the patient and the treating physician optimal treatment is the goal. Optimal treatment is a combination of many things, but first of all efficacy and minimal side effects. It would be helpful in choosing the right treatment for the individual patient if efficacy and side effects could, at least partly, be predicted by known genetic variants. For the scientist the interesting thing about the genetic background of response to hormone replacement therapy or other treatments is that it may provide information about the importance of a specific genes for bone health and about the functionality of the genetic variants.

Although some evidence suggests that the X allele of the *Xba*I polymorphism and the P allele of the *Pvu*II polymorphism in the estrogen receptor a gene, the long TTTA repeat allele of the *CYP19* gene along with polymorphisms in the TGF- β 1, *OPG* and *SOST* genes influence the response to postmenopausal hormone replacement therapy, we are still far from achieving these goals. The vast majority of the studies on the interaction between hormone replacement therapy and genetic variants are performed in too small populations resulting in severely underpowered statistical analyses with high risk of overlooking a true association or reporting chance findings. Most studies have less than 100 participants and the largest studies have less than 500 participants on long term hormone replacement therapy. Even 500 participants do not provide statistical power to perform complex analyses of gene – gene or gene – environmental interactions. Furthermore, the primary aims of many of the studies were not to investigate the interaction between hormone replacement therapy and genetic variants and some

of the studies therefore lack important information that could help understand the underlying mechanisms for example information about dietary intake, smoking and serum samples that would make it possible to perform additional analyses. A serious obstacle to change this is, that it has become impossible to conduct randomized trials on hormone replacement therapy with the knowledge we have on serious side effects to long-term estrogen treatment. There will always be women that choose hormone replacement treatment due to menopausal symptoms and it would therefore be possible to investigate the interaction between hormone replacement therapy and genetic variants in this group of women. However, women who choose estrogen treatment with all the information available about risk of breast cancer and cardiovascular disease are predominantly women with severe menopausal symptoms and we have to bear in mind that the severity of menopausal symptoms may be influenced by the same set of genetic variants as the ones we want to investigate or by other genetic variants or environmental factors. One way or the other this may lead to that any group of women, who have chosen hormone replacement therapy, constitutes a group of women with a specific genetic background and the results obtained may therefore not be applicable to the entire population of postmenopausal women. A better approach would therefore be for researchers who already have detailed information about hormone replacement therapy, bone phenotypes, covariates and genotypes from randomized study or high quality cohort studies to combine these data for further analyses.

References

1. Seeman E. Clinical review 137: Sexual dimorphism in skeletal size, density, and strength. *J Clin Endocrinol Metab.* 2001;86: 4576-4584.
2. Mosekilde L, Beck-Nielsen H, Sorensen OH, Nielsen SP, Charles P, Vestergaard P, Hermann AP, Gram J, Hansen TB, Abrahamson B, Ebbesen EN, Stilgren L, Jensen LB, Brot C, Hansen B, Tofteng CL, Eiken P, Kolthoff N. Hormonal replacement therapy reduces forearm fracture incidence in recent postmenopausal women - results of the Danish Osteoporosis Prevention Study. *Maturitas.* 2000;36:181-193.
3. Cauley JA, Robbins J, Chen Z, Cummings SR, Jackson RD, LaCroix AZ, LeBoff M, Lewis CE, McGowan J, Neuner J, Pettinger M, Stefanick ML, Wactawski-Wende J, Watts NB. Effects of estrogen plus progestin on risk of fracture and bone mineral density: the Women's Health Initiative randomized trial. *JAMA.* 2003;290: 1729-1738
4. de Bruin JP, Bovenhuis H, van Noord PA, Pearson PL, van Arendonk JA, te Velde ER, Kuurman WW, Dorland M. The role of genetic factors in age at natural menopause. *Hum Reprod.* 2001;16: 2014-2018.
5. Murabito JM, Yang Q, Fox C, Wilson PW, Cupples LA. Heritability of age at natural menopause in the Framingham Heart Study. *J Clin Endocrinol Metab.* 2005;90:3427-3430.
6. Gosden RG, Treloar SA, Martin NG, Cherkas LF, Spector TD, Faddy MJ, Silber SJ. Prevalence of premature ovarian failure in monozygotic and dizygotic twins. *Hum Reprod.* 2007;22:610-615.
7. Makovey J, Nguyen TV, Naganathan V, Wark JD, Sambrook PN. Genetic effects on bone loss in peri- and postmenopausal women: a longitudinal twin study. *J Bone Miner Res.* 2007;22:1773-1780.
8. Snieder H, MacGregor AJ, Spector TD. Genes control the cessation of a woman's reproductive life: a twin study of hysterectomy and age at menopause. *J Clin Endocrinol Metab.* 1998;83:1875-1880.
9. Locatelli I, Lichtenstein P, Yashin AI. The heritability of breast cancer: a Bayesian correlated frailty model applied to Swedish twins data. *Twin Res.* 2004;7:182-191.
10. Boyd NF, Dite GS, Stone J, Gunasekara A, English DR, McCredie

- MR, Giles GG, Trichler D, Chiarelli A, Yaffe MJ, Hopper JL. Heritability of mammographic density, a risk factor for breast cancer. *N Engl J Med.* 2002;347:886-894.
11. Mack TM, Hamilton AS, Press MF, Diep A, Rappaport EB. Heritable breast cancer in twins. *Br J Cancer.* 2002;87:294-300.
 12. del Senno L, Aguiari GL, Piva R. Dinucleotide repeat polymorphism in the human estrogen receptor (ESR) gene. *Hum Mol Genet.* 1992;1:354.
 13. Langdahl BL, Lokke E, Carstens M, Stenkjaer LL, Eriksen EF. A TA repeat polymorphism in the estrogen receptor gene is associated with osteoporotic fractures but polymorphisms in the first exon and intron are not (In Process Citation). *J Bone Miner Res.* 2000;15:2222-2230.
 14. van Meurs JB, Schuit SC, Weel AE, van der KM, Bergink AP, Arp PP, Colin EM, Fang Y, Hofman A, Van Duijn CM, van Leeuwen JP, Pols HA, Uitterlinden AG. Association of 5' estrogen receptor alpha gene polymorphisms with bone mineral density, vertebral bone area and fracture risk. *Hum Mol Genet.* 2003;12:1745-1754.
 15. Yim CH, Choi JT, Choi HA, Kang YS, Moon IG, Yoon HK, Han IK, Kang DH, Han KO. Association of estrogen receptor alpha gene microsatellite polymorphism with annual changes in bone mineral density in Korean women with hormone replacement therapy. *J Bone Miner Metab.* 2005;23:395-400.
 16. Castagnoli A, Maestri I, Bernardi F, del Senno L. PvuII RFLP inside the human estrogen receptor gene. *Nucleic Acids Res.* 1987;15:866.
 17. Andersen TI, Heimdal KR, Skrede M, Tveit K, Berg K, Borresen AL. Oestrogen receptor (ESR) polymorphisms and breast cancer susceptibility. *Hum Genet.* 1994;94:665-670.
 18. Kobayashi S, Inoue S, Hosoi T, Ouchi Y, Shiraki M, Orimo H. Association of bone mineral density with polymorphism of the estrogen receptor gene. *J Bone Miner Res.* 1996;11:306-311.
 19. Albagha OM, Pettersson U, Stewart A, McGuigan FE, MacDonald HM, Reid DM, Ralston SH. Association of oestrogen receptor alpha gene polymorphisms with postmenopausal bone loss, bone mass, and quantitative ultrasound properties of bone. *J Med Genet.* 2005;42:240-246.
 20. Ioannidis JP, Ralston SH, Bennett ST, Brandi ML, Grinberg D, Karassa FB, Langdahl B, van Meurs JB, Mosekilde L, Scollen S, Albagha OM, Bustamante M, Carey AH, Dunning AM, Enjuanes A, van Leeuwen JP, Mavilia C, Masi L, McGuigan FE, Nogues X, Pols HA, Reid DM, Schuit SC, Sherlock RE, Uitterlinden AG. Differential genetic effects of ESR1 gene polymorphisms on osteoporosis outcomes. *JAMA* 2004;292:2105-2114.
 21. Salmen T, Heikkinen AM, Mahonen A, Kroger H, Komulainen M, Saarikoski S, Honkanen R, Maenpaa PH. Early postmenopausal bone loss is associated with PvuII estrogen receptor gene polymorphism in Finnish women: effect of hormone replacement therapy. *J Bone Miner Res.* 2000;15:315-321.
 22. Deng HW, Li J, Li JL, Johnson M, Gong G, Davis KM, Recker RR. Change of bone mass in postmenopausal Caucasian women with and without hormone replacement therapy is associated with vitamin D receptor and estrogen receptor genotypes. *Hum Genet.* 1998;103:576-585.
 23. Sowers M, Jannausch ML, Liang W, Willing M. Estrogen receptor genotypes and their association with the 10-year changes in bone mineral density and osteocalcin concentrations. *J Clin Endocrinol Metab.* 2004;89:733-739.
 24. Rapuri PB, Gallagher JC, Knezetic JA, Haynatzka V. Estrogen receptor alpha gene polymorphisms are associated with changes in bone remodeling markers and treatment response to estrogen. *Maturitas.* 2006;53:371-379.
 25. Kobayashi N, Fujino T, Shirogane T, Furuta I, Kobamatsu Y, Yae-gashi M, Sakuragi N, Fujimoto S. Estrogen receptor alpha polymorphism as a genetic marker for bone loss, vertebral fractures and susceptibility to estrogen. *Maturitas.* 2002;41:193-201.
 26. Ongphiphadhanakul B, Chanprasertyothin S, Payatikul P, Tung SS, Piaseu N, Chailurkit L, Chansirikarn S, Puavilai G, Rajatanavin R. Oestrogen-receptor-alpha gene polymorphism affects response in bone mineral density to oestrogen in postmenopausal women. *Clin Endocrinol (Oxf).* 2000;52:581-585.
 27. Kurabayashi T, Tomita M, Matsushita H, Yahata T, Honda A, Takakuwa K, Tanaka K. Association of vitamin D and estrogen receptor gene polymorphism with the effect of hormone replacement therapy on bone mineral density in Japanese women. *Am J Obstet Gynecol.* 1999;180:1115-1120.
 28. Han KO, Moon IG, Kang YS, Chung HY, Min HK, Han IK. Nonassociation of estrogen receptor genotypes with bone mineral density and estrogen responsiveness to hormone replacement therapy in Korean postmenopausal women (see comments). *J Clin Endocrinol Metab.* 1997;82:991-995.
 29. Giguere Y, Dodin S, Blanchet C, Morgan K, Rousseau F. The association between heel ultrasound and hormone replacement therapy is modulated by a two-locus vitamin D and estrogen receptor genotype. *J Bone Miner Res.* 2000;15:1076-1084.
 30. Kurabayashi T, Matsushita H, Tomita M, Kato N, Kikuchi M, Nagata H, Honda A, Yahata T, Tanaka K. Association of vitamin D and estrogen receptor gene polymorphism with the effects of longterm hormone replacement therapy on bone mineral density. *J Bone Miner Metab.* 2004;22:241-247.
 31. Silvestri S, Thomsen AB, Gozzini A, Bagger Y, Christiansen C, Brandi ML. Estrogen receptor alpha and beta polymorphisms: is there an association with bone mineral density, plasma lipids, and response to postmenopausal hormone therapy? *Menopause.* 2006;13:451-461.
 32. Salmen T, Heikkinen AM, Mahonen A, Kroger H, Komulainen M, Saarikoski S, Honkanen R, Maenpaa PH. The protective effect of hormone-replacement therapy on fracture risk is modulated by estrogen receptor alpha genotype in early postmenopausal women. *J Bone Miner Res.* 2000;15:2479-2486.
 33. Weel AE, Uitterlinden AG, Westendorp IC, Burger H, Schuit SC, Hofman A, Helmerhorst TJ, van Leeuwen JP, Pols HA. Estrogen receptor polymorphism predicts the onset of natural and surgical menopause. *J Clin Endocrinol Metab.* 1999;84:3146-3150.
 34. Masi L, Becherini L, Gennari L, Amedei A, Colli E, Falchetti A, Farci M, Silvestri S, Gonnelli S, Brandi ML. Polymorphism of the aromatase gene in postmenopausal Italian women: distribution and correlation with bone mass and fracture risk. *J Clin Endocrinol Metab.* 2001;86:2263-2269.
 35. Gennari L, Masi L, Merlotti D, Picariello L, Falchetti A, Tanini A, Mavilia C, Del Monte F, Gonnelli S, Lucani B, Gennari C, Brandi ML. A polymorphic CYP19 TTTA repeat influences aromatase activity and estrogen levels in elderly men: effects on bone metabolism. *J Clin Endocrinol Metab.* 2004;89:2803-2810.
 36. Tofteng CL, Kindmark A, Brandstrom H, Abrahamson B, Petersen S, Stiger F, Stilgren LS, Jensen JE, Vestergaard P, Langdahl BL, Mosekilde L. Polymorphisms in the CYP19 and AR Genes-Relation to Bone Mass and Longitudinal Bone Changes in Postmenopausal Women With or Without Hormone Replacement Therapy: The Danish Osteoporosis Prevention Study. *Calcif Tissue Int.* 2003.
 37. Morrison NA, Yeoman R, Kelly PJ, Eisman JA. Contribution of trans-acting factor alleles to normal physiological variability: vitamin D receptor gene polymorphism and circulating osteocalcin. *Proc Natl Acad Sci U S A.* 1992;89:6665-6669.
 38. Baker AR, McDonnell DP, Hughes M, Crisp TM, Mangelsdorf DJ, Haussler MR, Pike JW, Shine J, O'Malley BW. Cloning and expression of full-length cDNA encoding human vitamin D receptor. *Proc Natl Acad Sci U S A.* 1988;85:3294-3298.
 39. Ingles SA, Haile RW, Henderson BE, Kolonel LN, Nakaichi G, Shi CY, Yu MC, Ross RK, Coetzee GA. Strength of linkage disequilibrium between two vitamin D receptor markers in five ethnic groups: implications for association studies. *Cancer Epidemiol Biomarkers Prev.* 1997;6:93-98.
 40. Durrin LK, Haile RW, Ingles SA, Coetzee GA. Vitamin D receptor 3'-untranslated region polymorphisms: lack of effect on mRNA stability. *Biochim Biophys Acta* 1999;1453:311-320.
 41. Cooper GS, Umbach DM. Are vitamin D receptor polymorphisms associated with bone mineral density? A meta-analysis (see comments). *J Bone Miner Res.* 1996;11:1841-1849.
 42. Gong G, Stern HS, Cheng SC, Fong N, Mordeson J, Deng HW, Recker RR. The association of bone mineral density with vita-

- min D receptor gene polymorphisms. *Osteoporos Int.* 1999;9:55-64.
43. Uitterlinden AG, Ralston SH, Brandi ML, Carey AH, Grinberg D, Langdahl BL, Lips P, Lorenc R, Obermayer-Pietsch B, Reeve J, Reid DM, Amidei A, Bussiti A, Bustamante M, Husted LB, Diez-Perez A, Dobnig H, Dunning AM, Enjuanes A, Fahrleitner-Pammer A, Fang Y, Karczmarewicz E, Kruk M, van Leeuwen JP, Mavilia C, van Meurs JB, Mangion J, McGuigan FE, Pols HA, Renner W, Rivadeneira F, van Schoor NM, Scollen S, Sherlock RE, Ioannidis JP. The association between common vitamin D receptor gene variations and osteoporosis: a participant-level meta-analysis. *Ann Intern Med.* 2006;145:255-264.
 44. Grant SF, Reid DM, Blake G, Herd R, Fogelman I, Ralston SH. Reduced bone density and osteoporosis associated with a polymorphic Sp1 binding site in the collagen type I alpha 1 gene. *Nat Genet.* 1996;14:203-205.
 45. Langdahl BL, Ralston SH, Grant SF, Eriksen EF. An Sp1 binding site polymorphism in the COL1A1 gene predicts osteoporotic fractures in both men and women. *J Bone Miner Res.* 1998;13:1384-1389.
 46. Uitterlinden AG, Burger H, Huang Q, Yue F, McGuigan FE, Grant SF, Hofman A, van Leeuwen JP, Pols HA, Ralston SH. Relation of alleles of the collagen type I alpha 1 gene to bone density and the risk of osteoporotic fractures in postmenopausal women (see comments). *N Engl J Med.* 1998;338:1016-1021.
 47. Mann V, Hobson EE, Li B, Stewart TL, Grant SF, Robins SP, Aspden RM, Ralston SH. A COL1A1 Sp1 binding site polymorphism predisposes to osteoporotic fracture by affecting bone density and quality. *J Clin Invest.* 2001;107:899-907.
 48. Efstathiadou Z, Tsatsoulis A, Ioannidis JP. Association of collagen alpha 1 Sp1 polymorphism with the risk of prevalent fractures: a meta-analysis. *J Bone Miner Res.* 2001;16:1586-1592.
 49. Ralston SH, Uitterlinden AG, Brandi ML, Balcells S, Langdahl BL, Lips P, Lorenc R, Obermayer-Pietsch B, Scollen S, Bustamante M, Husted LB, Carey AH, Diez-Perez A, Dunning AM, Falchetti A, Karczmarewicz E, Kruk M, van Leeuwen JP, van Meurs JB, Mangion J, McGuigan FE, Mellibovsky L, Del Monte F, Pols HA, Reeve J, Reid DM, Renner W, Rivadeneira F, van Schoor NM, Sherlock RE, Ioannidis JP. Large-scale evidence for the effect of the COL1A1 Sp1 polymorphism on osteoporosis outcomes: the GENOMOS study. *PLoS Med.* 2006;3:e90.
 50. MacDonald HM, McGuigan FA, New SA, Campbell MK, Golden MH, Ralston SH, Reid DM. COL1A1 Sp1 polymorphism predicts perimenopausal and early postmenopausal spinal bone loss. *J Bone Miner Res.* 2001;16:1634-1641.
 51. Simsek M, Cetin Z, Bilgen T, Taskin O, Luleci G, Keser I. Effects of hormone replacement therapy on bone mineral density in Turkish patients with or without COL1A1 Sp1 binding site polymorphism. *J Obstet Gynaecol Res.* 2008;34:73-77.
 52. Gonzalez-Bofill N, Husted LB, Vestergaard P, Tofteng CL, Abrahamsen B, Eiken P, Langdahl BL. Effects of COL1A1 Polymorphisms and Haplotypes on Perimenopausal Bone mass, Postmenopausal Bone Loss and Effect of Hormone Replacement Therapy. In Preparation.
 53. Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, Boers GJ, den Heijer M, Kluijtmans LA, van den Heuvel LP. A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. *Nat Genet.* 1995;10:111-113.
 54. Lubec B, Fang-Kircher S, Lubec T, Blom HJ, Boers GH. Evidence for McKusick's hypothesis of deficient collagen cross-linking in patients with homocystinuria. *Biochim Biophys Acta.* 1996;1315:159-162.
 55. Miyao M, Morita H, Hosoi T, Kurihara H, Inoue S, Hoshino S, Shiraki M, Yazaki Y, Ouchi Y. Association of methylenetetrahydrofolate reductase (MTHFR) polymorphism with bone mineral density in postmenopausal Japanese women. *Calcif Tissue Int.* 2000;66:190-194.
 56. Abrahamsen B, Madsen JS, Tofteng CL, Stilgren L, Bladbjerg EM, Kristensen SR, Brixen K, Mosekilde L. A common methylenetetrahydrofolate reductase (C677T) polymorphism is associated with low bone mineral density and increased fracture incidence after menopause: longitudinal data from the Danish osteoporosis prevention study. *J Bone Miner Res.* 2003;18:723-729.
 57. Villadsen MM, Bunger MH, Carstens M, Stenkaer L, Langdahl BL. The methylenetetrahydrofolate reductase (MTHFR) gene polymorphism (C677T) is a weak predictor of BMD and associated with osteoporotic fractures. *Osteoporos Int.* 2005;16:411-416.
 58. Cambien F, Ricard S, Troesch A, Mallet C, Generenaz L, Evans A, Arveiler D, Luc G, Ruidavets JB, Poirier O. Polymorphisms of the transforming growth factor-beta 1 gene in relation to myocardial infarction and blood pressure. The Etude Cas-Temoin de l'Infarctus du Myocarde (ECTIM) Study (see comments). *Hypertension.* 1996;28:881-887.
 59. Awad MR, El Gamel A, Hasleton P, Turner DM, Sinnott PJ, Hutchinson IV. Genotypic variation in the transforming growth factor-beta1 gene: association with transforming growth factor-beta1 production, fibrotic lung disease, and graft fibrosis after lung transplantation. *Transplantation.* 1998;66:1014-1020.
 60. Langdahl BL, Carstens M, Stenkaer L, Eriksen EF. Polymorphisms in the transforming growth factor beta1 gene and osteoporosis. *Bone.* 2003;32:297-310.
 61. Keen RW, Snieder H, Molloy H, Daniels J, Chiano M, Gibson F, Fairbairn L, Smith P, MacGregor AJ, Gewert D, Spector TD. Evidence of association and linkage disequilibrium between a novel polymorphism in the transforming growth factor beta 1 gene and hip bone mineral density: a study of female twins. *Rheumatology (Oxford).* 2001;40:48-54.
 62. Yamada Y, Miyauchi A, Takagi Y, Tanaka M, Mizuno M, Harada A. Association of the C-509-->T polymorphism, alone or in combination with the T869-->C polymorphism, of the transforming growth factor-beta1 gene with bone mineral density and genetic susceptibility to osteoporosis in Japanese women. *J Mol Med.* 2001;79:149-156.
 63. Yamada Y, Miyauchi A, Goto J, Takagi Y, Okuizumi H, Kanematsu M, Hase M, Takai H, Harada A, Ikeda K. Association of a polymorphism of the transforming growth factor-beta1 gene with genetic susceptibility to osteoporosis in postmenopausal Japanese women. *J Bone Miner Res.* 1998;13:1569-1576.
 64. Bustamante M, Nogues X, Enjuanes A, Elosua R, Garcia-Giralt N, Perez-Edo L, Caceres E, Carreras R, Mellibovsky L, Balcells S, Diez-Perez A, Grinberg D. COL1A1, ESR1, VDR and TGFB1 polymorphisms and haplotypes in relation to BMD in Spanish postmenopausal women. *Osteoporos Int.* 2007;18:235-243.
 65. Langdahl BL, Uitterlinden AG, Ralston SH, Trikalinos TA, Balcells S, Brandi ML. Large-scale analysis of association between polymorphisms in the Transforming Growth Factor Beta 1 gene (TGFB1) and osteoporosis: The GENOMOS Study. *Bone.* In print.
 66. Langdahl BL, Abrahamsen B, Vestergaard P, Tofteng CL, Kolthoff N, Madsen JS, Mosekilde L. Polymorphisms in the Transforming Growth Factor Beta1 gene and Perimenopausal Bone Mass, Early Postmenopausal Bone Loss and Response to Hormone Replacement Therapy. *Calcif Tissue Int.* Abstract 2003.
 67. Simonet WS, Lacey DL, Dunstan CR, Kelley M, Chang MS, Luthy R, Nguyen HQ, Wooden S, Bennett L, Boone T, Shimamoto G, DeRose M, Elliott R, Colombero A, Tan HL, Trail G, Sullivan J, Davy E, Bucay N, Renshaw-Gegg L, Hughes TMR, Hill D, Pattison W, Campbell P, Boyle WJ. Osteoprotegerin: a novel secreted protein involved in the regulation of bone density [see comments]. *Cell.* 1997;89:309-319.
 68. Langdahl BL, Carstens M, Stenkaer L, Eriksen EF. Polymorphisms in the osteoprotegerin gene are associated with osteoporotic fractures. *J Bone Miner Res.* 2002;17:1245-1255.
 69. Arko B, Prezelj J, Komel R, Kocijancic A, Hudler P, Marc J. Sequence variations in the osteoprotegerin gene promoter in patients with postmenopausal osteoporosis. *J Clin Endocrinol Metab.* 2002;87:4080-4084.
 70. Brandstrom H, Stiger F, Lind L, Kahan T, Melhus H, Kindmark A. A single nucleotide polymorphism in the promoter region of the human gene for osteoprotegerin is related to vascular morphology and function. *Biochem Biophys Res Commun.* 2002;293:13-17.
 71. Richards JB, Rivadeneira F, Inouye M, Pastinen TM, Soranzo N,

- Wilson SG, Andrew T, Falchi M, Gwilliam R, Ahmadi KR, Valdes AM, Arp P, Whittaker P, Verlaan DJ, Jhamai M, Kumanduri V, Moorhouse M, van Meurs JB, Hofman A, Pols HA, Hart D, Zhai G, Kato BS, Mullin BH, Zhang F, Deloukas P, Uitterlinden AG, Spector TD. Bone mineral density, osteoporosis, and osteoporotic fractures: a genome-wide association study. *Lancet*. 2008;371:1505-1512.
72. Langdahl BL, Vestergaard P, Abrahamsen B, Tofteng CL, Kolthoff N, Mosekilde L. Polymorphisms in the Promoter of the Osteoprotegerin Gene are associated with reduced Perimenopausal Bone Mass. *J Bone Miner Res*. Abstract 2002.
73. Little RD, Carulli JP, Del Mastro RG, Dupuis J, Osborne M, Folz C, Manning SP, Swain PM, Zhao SC, Eustace B, Lappe MM, Spitzer L, Zweier S, Braunschweiger K, Benchekroun Y, Hu X, Adair R, Chee L, FitzGerald MG, Tulig C, Caruso A, Tzellas N, Bawa A, Franklin B, McGuire S, Noguez X, Gong G, Allen KM, Anisowicz A, Morales AJ, Lomedico PT, Recker SM, Van Eerdewegh P, Recker RR, Johnson ML. A mutation in the LDL receptor-related protein 5 gene results in the autosomal dominant high-bone-mass trait. *Am J Hum Genet*. 2002;70:11-19.
74. Gong Y, Slee RB, Fukai N, Rawadi G, Roman-Roman S, Reginato AM, Wang H, Cundy T, Glorieux FH, Lev D, Zacharin M, Oexle K, Marcelino J, Suwairi W, Heeger S, Sabatakos G, Apte S, Adkins WN, Allgrove J, Arslan-Kirchner M, Batch JA, Beighton P, Black GC, Boles RG, Boon LM, Borrone C, Brunner HG, Carle GF, Dalapiccola B, De Paepe A, Floege B, Halfhide ML, Hall B, Hennekam RC, Hirose T, Jans A, Juppner H, Kim CA, Keppler-Noreuil K, Kohlschuetter A, LaCombe D, Lambert M, Lemyre E, Letteboer T, Peltonen L, Ramesar RS, Romanengo M, Somer H, Steichen-Gersdorf E, Steinmann B, Sullivan B, Superti-Furga A, Swoboda W, van den Boogaard MJ, Van Hul W, Vikkula M, Votruba M, Zabel B, Garcia T, Baron R, Olsen BR, Warman ML. LDL receptor-related protein 5 (LRP5) affects bone accrual and eye development. *Cell*. 2001;107:513-523.
75. van Meurs JB, Trikalinos TA, Ralston SH, Balcells S, Brandi ML, Brixen K, Kiel DP, Langdahl BL, Lips P, Ljunggren O, Lorenc R, Obermayer-Pietsch B, Ohlsson C, Pettersson U, Reid DM, Rousseau F, Scollen S, Van Hul W, Agueda L, Akesson K, Benevolenskaya LI, Ferrari SL, Hallmans G, Hofman A, Husted LB, Kruk M, Kaptoge S, Karasik D, Karlsson MK, Lorentzon M, Masi L, McGuigan FE, Mellstrom D, Mosekilde L, Noguez X, Pols HA, Reeve J, Renner W, Rivadeneira F, van Schoor NM, Weber K, Ioannidis JP, Uitterlinden AG. Large-scale analysis of association between LRP5 and LRP6 variants and osteoporosis. *JAMA*. 2008;299:1277-1290.
76. Gonzalez-Bofill N, Husted LB, Vestergaard P, Tofteng CL, Abrahamsen B, Eiken P, Langdahl BL. LRP5 and LRP6 Polymorphisms Affect Peak Bone Mass but Not Early Postmenopausal Bone Loss. *J Bone Miner Res*. Abstract 2006.
77. Brunkow ME, Gardner JC, Van Ness J, Paepfer BW, Kovacevich BR, Proll S, Skonier JE, Zhao L, Sabo PJ, Fu Y, Alisch RS, Gillett L, Colbert T, Tacconi P, Galas D, Hamersma H, Beighton P, Mulligan J. Bone dysplasia sclerosteosis results from loss of the SOST gene product, a novel cystine knot-containing protein. *Am J Hum Genet*. 2001;68:577-589.
78. Balemans W, Patel N, Ebeling M, Van Hul E, Wuyts W, Laca C, Dioszegi M, Dikkers FG, Hildering P, Willems PJ, Verheij JB, Lindpaintner K, Vickery B, Foerzler D, Van Hul W. Identification of a 52 kb deletion downstream of the SOST gene in patients with van Buchem disease. *J Med Genet*. 2002;39:91-97.
79. Uitterlinden AG, Arp PP, Paepfer BW, Charmley P, Proll S, Rivadeneira F, Fang Y, van Meurs JB, Britschgi TB, Latham JA, Schatzman RC, Pols HA, Brunkow ME. Polymorphisms in the sclerosteosis/van Buchem disease gene (SOST) region are associated with bone-mineral density in elderly whites. *Am J Hum Genet*. 2004;75:1032-1045.
80. Gonzalez-Bofill N, Husted LB, Vestergaard P, Tofteng CL, Abrahamsen B, Eiken P, Langdahl BL. SRP3 and SRP9 Polymorphisms in the SOST Gene are Associated with Perimenopausal Bone Mass and Early Postmenopausal Bone Loss. *Calcif Tissue Int*. Abstract 2007.