

# Bone regeneration in dentistry

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## Summary

**The edentulism of the jaws and the periodontal disease represent conditions that frequently leads to disruption of the alveolar bone. The loss of the tooth and of its bone of support lead to the creation of crestal defects or situation of maxillary atrophy. The restoration of a functional condition involves the use of endosseous implants who require adequate bone volume, to deal with the masticatory load. In such situations the bone need to be regenerated, taking advantage of the biological principles of osteogenesis, osteoinduction and osteoconduction. Several techniques combine these principles with different results, due to the condition of the bone base on which we operate changes, the surgical technique that we use, and finally for the bone metabolic conditions of the patient who can be in a state of systemic osteopenia or osteoporosis; these can also affect the result of jaw bone reconstruction.**

*KEY WORDS: osteoporosis; edentulism; guided bone regeneration.*

## Introduction

The loss of the tooth causes the resorption of the alveolar bone (1). As to stop the stimulus induced by the periodontal ligament, the vestibular cortical bone is subjected to resorption and the marrow component of the alveolous gradually disappeared (2-4). The consequence is the change of the morphology of the alveolar ridge, which, in limited form for number of teeth lost, configure the degree of the alveolar defect and by extension, condition of more pronounced atrophy (5). The gradual disappearance of the alveolar process involves the reduction of sagittal size and then vertical size of the

jaws, as described firstly by the classifications of Cawood and Howell (6) and then by Misch and Judy (7) leading to a subversion of intermaxillary relations and functional abnormality which makes incompetent the two dental arches.

Preserving as restoring a sufficient bone volume to support the prosthetic load, and also the insertion of the dental implant as a support for prosthesis, requires the use of surgical protocols that enable the bone regeneration on the deficient sites, using the principles of osteogenesis, osteoinduction and osteoconduction.

The jaw bone will respond to these protocols in a very subjective way because of the bone site to be restored, of the operative protocol and the general bone conditions which are sometimes deficiency because of osteopenia or osteoporosis.

## Aims

The different conditions of decreased bone jaw can be corrected with clinical protocols associating biological principles of bone regeneration. However, the volume of regenerated bone has to support the masticatory forces transmitted by the implant, with different results for each jaw and for patient's different metabolic conditions. These will be briefly considered.

## Biological Principles

Osteogenesis, osteoinduction and osteoconduction are the biological principles that offer the possibility to regenerate lost bone volume.

The first one allows the use of autologous bone: osteoblastic cells and Haversian systems of the grafted bone fragment will be replaced by newly formed bone from the walls of the recipient bed (8). The osteoinduction enables migrations and proliferation of connective undifferentiated cells in the site to be regenerated. This potential differentiation is conditioned by the presence of growth factors (GF) on the site (9).

The osteoconduction is the ability of a material to operate as a scaffold to guide the tissue regeneration. The material will also partially be replaced by newly formed cells (10).

Several techniques allow the application of these principles. The results change for quantity and quality and depending on the type of principle that is used. In fact, within the jaw bone can be exploited: the repair, guided repair and regeneration.

The repair is the formation in a bone defect of a part of connective tissue formed by cells and fibroblasts, which in part will be replaced by osteoblasts that will deposit an osteoid matrix that will ossify (11). However, the volume of regenerated tissue will be lower than expected (12) for the interference of non osteocompetent cells.

The guided repair uses the principle of resorption/substitution of an osteointegrated biomaterial with new-bone. The result will depend on the features of osteoconductive grafted material and provide a tissue in which tracks of the same will long remain (13). The regeneration is limited to the implementation of undifferentiated connective cells present in the site to be regenerated by appropriate clinical solution which isolate the site (14), or to the bone formation obtained from autologous vital material inserted into the defect.

## Operative protocols

Several techniques allow restoring lost alveolar bone with formation of stable bone matrix (Table 1).

**Guided Bone Regeneration (GBR):** it allows, through the use of resorbable or non-resorbable membrane, the filling of a defect through the guided growth of only osteogenetic strains and preventing the invasion of non-osteogenetic tissues which are competitive with the bone itself (15, 16). Among the devices used to isolate the defect, in addition to the membranes, we have also the grids that allow you to keep the space needed for bone formation avoiding the collapse of soft tissue (17). The widespread use of resorbable membranes free of mechanical consistency has meant that these defects will be filled from osseointegrative biomaterials used as support (18). **Use of growth factors (GF):** these are glycoproteins with autocrine and paracrine function that grafted in the site, recruit and multiply the osteocompetent cellular strains. For the clinical use the platelet-rich plasma (PRP) (19, 20) and the platelet rich fibrin (PRF) (21) are used as autologous materials obtained from the patient's blood by centrifugation (22); we can use also bone morphogenetic proteins (BMPs) obtained by genetic engineering (OP-1). All these factors may be included alone or together with a biocompatible material acting as support (23), so that their action is prolonged for a few days. We can distinguish between the graft materials: autologous graft (autograft), homologous graft (allograft), heterologous graft (xenograft) and alloplastic graft.

The autologous bone is the gold standard because it contains the three properties. Depending on the size of the defect is used to harvest intraoral or extraoral. The grafted material maintains the characteristics of embryological site of origin: bone density, which matures on the site reflects that principle (24).

The allograft is provided by the tissue banks in various formulations as sticks, granules or paste. This is an osteoconductive material that provides mechanical properties even in large defects (25).

The heterologous material, that has bovine or equine origin, is a non-stoichiometric apatite less resorbable, which does not resist to the traction forces and to the masticatory load (26, 27).

The alloplastic are osteointegrative materials with a different degree of resorption; they have biomechanical properties; they are partially replaced in bone remodeling based on their size and porosity (28). The surgical procedures for the grafting materials are: the fixation of graft of sticks or bone particulate rigidly to the atrophic base or the filling of defects with bone particles. For defects of the superior alveolar process we can use the mini or large maxillary sinus lift with the crestal or lateral surgical approach and for the sagittal deficiency of the site we can use the distraction osteogenesis.

The aim of the bone regeneration is to insert the titanium implant in its context. This alloplastic insert, whose rough and porous surface (29) allows osteointegration with the bone tissue (30, 31) and it will provide the prosthetic support solution for the clinical case.

## The bone metabolism and the atrophy of the jaws

These protocols should take into consideration the patient's osseous metabolic condition. In fact, there are systematic conditions of osteopenia and osteoporosis which may also be reflected in the maxillary area (32). These diseases can be linked to regressive states (post-menopausal, senile) or secondary to osteomalacia, hyperparathyroidism, disendocrinopathy, metabolic disorders (33).

These clinical disorders include preservation of bone mass but in the marrow and in the cortical component we can note a less production of the osteoid matrix, a slow mineralization of the same, a trend accelerated remodeling with fracture of the trabeculae of less caliber (34).

The diagnosis of these conditions is not easy with the radiological diagnostic equipment as intraoral-radiograph and panoramic radiograph, even if you can get more information with Computer Tomography (CT).

While the biochemical investigation is useful both in the initial assessment of patients undergoing these treatment programs, both in advanced stages of investigation for more complex cases.

There are several tests that, in case of suspected osteoporosis, you can make in the first phase or, when the information obtained from these initial biochemical investigations are not conclusive, in the second phase (Table 2) (35).

With regard to maxillary osteoporosis most of the authors agree that the skeletal osteoporosis and the maxillary osteoporosis are often associated (36) but we don't exclude situations where there are general conditions of osteoporosis, osteopenia, that don't involve the bone level of the jaws impact and on the contrary. Taguchi (76) notes the correlation between mandibular bone resorption and decreased vertebral bone density; Drage (77) by analyzing densitometry vertebrae, femurs and maxillary and comparing them, find only correlation between the mandible and the femur or vertebrae.

## Discussion

Currently Oral Surgeon is in possession of numerous instruments which, using basic biological principles, allow adequate jaw bone volume rehabilitation to insert an osseointegrated implant, able to support a prosthetic restoration.

In literature, regenerative protocols techniques are associated with a high number of complications (37).

Using a GBR technique, the main complication is flap dehiscence with infection of the membrane and the grafted material. Jensen (38) records the need to second surgery to obtain sufficient bone volume in a percentage of cases ranging between 4.1% and 32%.

The use of resorbable membranes and techniques of horizontal regeneration have got fewer complications (39).

In autologous bone grafts the main complication of the receiving site is always the flap dehiscence associated with graft infection (40), but in these cases we also have to consider the donor site complications. Grafts more associated to post-operative problems are those from iliac crest and chin, while the less ones are those from mandibular ramus and calvaria crest. Although calvaria graft provides an optimal bone quantity and quality, it is difficult to be accepted by patients (41, 42).

Surgeon who intends to approach to these regeneration techniques, certainly must know the implant survival rates in regenerated bone. Several systematic reviews show with GBR technique, used in vertical and/or horizontal augmentations, an implant survival rate of > 90% (43-47). These studies, however, do not consider numerous technical variables: it is then necessary to design new studies to assess factors related to the site and the individuality of the patient in considering the effectiveness and predictability of the GBR (75). For autologous bone grafts, the implant survival rate varies between 76% and 100% (46) with worse results for iliac crest bone compared to calvaria bone or intra-oral grafts (48).

However, given the multitude of techniques and materials existing, based on our current state of knowledge and on data from the literature, we could assess that there is not scientific evidence to indicate which technique is better (37, 38).

In choosing, clinician and patient must weigh the pros and cons based on what are biological and economic costs, and priority should be given to less invasive techniques, with fewer complications and reduced treatment time (48).

Failing to reach firm conclusions, regardless of the technique used to obtain predictable results, it is essential to respect some well established principles. Among these are fundamental: stability of the grafted material, primary closure of the flap, the angiogenesis to ensure the supply of undifferentiated mesenchymal cells (49).

All the regeneration techniques are affected by the bone area in which are carried out, ensuring that clinical outcome will be different by jaws area. In fact, blood supply of the grafted material is influenced

Table 1 - Type of maxillary and mandibular defect and the bone regeneration technique.

Type of defect	Maxillary		Mandibular	
Width Reduction	Good bone density: GBR or split crest and implant at the same time	Poor bone density: GBR or split crest or graft and implant after a period of healing	Good bone density: GBR or split crest and implant at the same time	Poor bone density: GBR or split crest or graft and implant after a period of healing
Width and height Reduction	Three clinical situation: 1) height and width sufficient to insert an implant 2) pneumatization of maxillary sinus not associated with alveolar bone resorption: lateral or crestal sinus lift 3) pneumatization of maxillary sinus associated with alveolar bone resorption: sinus lift and onlay graft		Two clinical situation: 1) height and width sufficient to insert a short implant 2) Regeneration (GBR, distraction osteogenesis, block graft) and after a period of healing implant insertion	

Table 2 - Different type of test to investigate osteoporosis in initial clinical phase and in the second phase.

Initial biochemical investigations	In-depth investigations
<ul style="list-style-type: none"> <li>- ESR</li> <li>- CBC</li> <li>- Serum Protein Electrophoresis</li> <li>- Serum-calcium levels</li> <li>- Phosphorus</li> <li>- Total alkaline phosphatase</li> <li>- Creatinine</li> <li>- calciuria 24-hour</li> </ul>	<ul style="list-style-type: none"> <li>- Ionized calcium</li> <li>- PTH</li> <li>- 25-hydroxyvitamin D serum</li> <li>- Specific hormones (TSH, cortisoluria in 24 hours, testosterone for men)</li> <li>- Antibodies anti-amigdala, endomysial and antitransglutaminasi</li> <li>- Protein immunofixation, serum and urinary</li> <li>- Serum tryptase, urinary N-methylhistamine</li> <li>- Specific markers of bone turnover and sensitive</li> <li>- Bone marrow aspirate</li> <li>- Bone biopsy</li> </ul>

by site-specific location of the overall bone marrow, which is more sensitive to regeneration because is more vascularised than the cortical one, less disposed to metabolic exchange.

Misch and Zarb (50) have classified jaw bone density by dividing cortical and medullary quote in different portions of mandibular and maxillary bones. Therefore, bone type D1 (cortical thick) is found in symphysis region; bone type D2 (thick cortical bone and thick medullary bone) in mandibular ramus; bone type D3 (thin and porous cortical bone and thin medullary bone) across the maxillary arch; bone type D4 (thin and large trabeculae) in the tuber maxillae.

The possibility of integration of a graft material in addition to the density parameter depends on the morphology of the residual ridge. This morphology from a clinical point of view influences the depth of the vestibule, the tension of the flap and thus the stability of the material after the suture (51). The severe reabsorbed edentulous mandibular ridge has got all these characteristics in the negative, in contrast to the maxillary areas.

Nissan (52) uses to rehabilitate posterior mandibular areas the bloc grafts fixed with mini screws and protected by a membrane and shows a grafting success rate of 79.3%. Sbordone (53) shows a resorption of onlay iliac bone graft in block of 42% if placed in the anterior maxillae and 59% when placed in the posterior areas of lower jaw. Calvaria bone graft, instead, is less affected by remodeling phenomena. Smolka et al. (54) reported at one year a graft volume reduction of 19.2%.

Keith (55), in dealing with 82 defects, gets a failure rate of 71% with

dehiscence and infections in the posterior lower jaw using homologous bone grafts in block. Things seem to go better in the maxilla: Ferri (56) using onlay autologous grafts reports an implant success rate of 97%, and he does not report phenomena of site infection, but complain as a major problem the graft resorption.

It could be concluded that the maxillary sites are more receptive to regenerative therapy especially when consider grafting material in block rather than in particulates, that could be explained by the lower blood supply of the atrophic mandibular edentulous ridge (57).

The porous bone while allowing a greater blood supply, promotes the regeneration techniques because it ensures a better trophism of the grafted material; at the same time being less dense it has got the worst mechanical properties and it suffers more the loads transmitted by prosthetic implant (58).

The guarantee of sufficient bone quantity and of a high bone density is a prerequisite to the biomechanical stability and implant osseointegration to maintain over time (59, 60).

A key role in maintaining bone grafted volume is played, however, by the implant: its active surface is the basis of the metabolic exchange processes with bone cells and growth factors that ensure the functioning of the bone / implant / prosthesis system.

Particularly important is the correct timing of implant surgery: in fact, drilling regenerated bone after bone grafting, to place the implant, will promote the disposal of Growth Factors behind the surface that will be in contact with the insert with a larger proportion of Bone Implant Contact (BIC) (61).



It is essential, in case we graft a biomaterial, including long waiting times until could be generated a part of mature vital bone. The type of biomaterial used affects the maturation of the regenerated tissue: in the case of autologous bone chips 3-4 months are sufficient for a 30% vital bone mineralization; in the case of alloplastic material and of homologous bone particles are needed even more times (62,78,79).

Waiting time, however, is indicative and may vary from subject to subject, and in different sites in the same subject especially in the presence of osteopenia or osteoporosis, situations in which the bone metabolism is altered and the formation of vital material will be delayed.

These diseases are not absolute contraindications for the regeneration techniques (63) and the subsequent implant therapy (64), even if they reported a higher percentage of failures and complications. Naturally, modifiable risk factors for osteoporosis should be removed, patient lifestyle should be changed and secondary forms of this disease should be treated (63).

Currently, a point of particular attention is the possibility, through systemic and / or local interventions, to promote the mineralization of regenerated bone by recruiting Vitamin D and Calcium in adequate doses, as expressed by Cooper in 1998 (65).

There are also studies on animal model showing that the administration of Bisphosphonates such as strontium ranelate, improves implant stability (66-68, 70). Let us note however that Bisphosphonates have been associated with osteonecrosis of the jaw (69) and before undertaking any therapy, surgical risks should be carefully assessed.

On the same animal model has been shown that treatment with calcitonin (71) or simvastatin (73) increases the amount of newly formed bone in defects treated with e-PTFE membranes, although statins according to the mode of administration and dosage, the effects can be void or against (74). Furthermore, Hormone Replacing Therapy seems to prevent the influence that estrogen deficiency exerts on bone healing in rats without ovary (72). But these are preliminary results in animal models that need further investigation in order to begin testing on humans *in vivo*.

## Conclusion

Regenerative therapy of atrophic edentulous maxilla is configured as a real social problem because of the importance of implant-prosthetic therapy. Lack to assess the metabolic conditions of the patient and its individual parameters is certainly a source of failures.

## References

1. Atwood DA. Reduction of residual ridge: A major oral disease entity. *J Prosthet Dent* 1971;26(3):267-279.
2. Neufeld JO. Changes in the trabecular pattern of the mandible following the loss of teeth. *J Prosthet Dent* 1958;8(4):685-697.
3. Ulm C, Solar P, Blahout R et al. Reduction of the compact and cancellous bone substances of the edentulous mandible caused by resorption. *Oral Surg Oral Med Oral Pathol* 1992;74:131-6.
4. Botticelli D, Berglundh T, Lindhe J. Hard tissue alterations following immediate implant placement in extraction sites. *J Clinical Periodontol* 2004;31(10):820-8.
5. Cho JY. The periodontist and the edentulous area-localised ridge augmentation. *Int Dent J* 1998;48(3 Suppl1):326-9.
6. Cawood JI, Howell RA. A classification of the edentulous jaws. *Int J Oral Maxillofac Surg* 1988;17:232-236.
7. Misch CE. Suddivisione dell'osso disponibile. In: *L'odontoiatria implantare contemporanea*. I edizione italiana Antonio Delfino Editore 2000 pag. 89-108.
8. Burchardt H. The biology of bone graft repair. *Clin Orthop Relat Res* 1983;174:28-42.
9. Sykaras N, Opperman LA. Bone morphogenetic proteins (BMPs): how

do they function and what can they offer the clinician? *Journal of Oral Science* 2003;45(2):57-73.

10. Albrektsson T, Johansson C. Osteoinduction, osteoconduction and osteointegration. *Eur Spine J* 2001;10:s96-s101.
11. Cardaropoli G, Araújo M, Lindhe J. Dynamics of bone tissue formation in tooth extraction sites. An experimental study in dogs. *J Clin Periodontol* 2003;30(9):809-818.
12. Schropp L, Wenzel A, Kostopoulos L et al. Bone healing and soft tissue contour changes following single-tooth extraction: a clinical and radiographic 12-month prospective study. *Int J Periodontics restorative dent.* 2003;23:313-323.
13. Lynch SE, Marx RE. Tissue engineering Grafting materials in repair and restoration. Miami Quintessence Books 2000:84-97.
14. Buser D, Bragger U, Lang NP et al. Regeneration and enlargement of jaw bone using guided tissue regeneration. *Clin Oral Impl Res* 1990;1(1):22-32.
15. Lang NP, Hammerle CHF, Bragger U et al. Guided tissue regeneration in jawbone defects prior to implant placement. *Clin Oral Impl Res* 1994;5(2):92-97.
16. Dahlin C, Linde A, Gottlow J. et al. Healing of bone defects by guided tissue regeneration. *Plastic Reconstr Surg* 1988;81(5):672-676.
17. Rocuzzo M, Ramieri G, Bunino M et al. Autogenous bone graft alone or associated with titanium mesh for vertical alveolar ridge augmentation: a controlled clinical trial. *Clin Oral Impl Res* 2007;18:286-294.
18. Zitzmann NU, Naef R, Schärer P. Resorbable Versus Nonresorbable Membranes in Combination With Bio-Oss for Guided Bone Regeneration. *Int J Oral Maxillofac Implants* 1997;12(6):844-852.
19. Anitua E. Plasma rich in growth factors: preliminary results of use in the preparation of future sites for implants. *Int J Oral Maxillofac Implants* 1999;14(4):529-535.
20. Tonelli P, Mannelli D, Brancato L et al. Titolazione del Platelet Derived Growth Factor e del Trasforming Growth Factor- $\beta$  nel Plasma ricco di piastrine, impiegato quale amplificatore della rigenerazione ossea mascellare. *Minerva Stomatologica* 2005;54(1-2):23-34.
21. Choukroun J, Diss A, Simonpieri A et al. Platelet-rich fibrin (PRF): A second-generation platelet concentrate. Part IV: Clinical effects on tissue healing. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006;101(3):e56-60.
22. Dohan Ehrenfest DM, Rasmusson L, Albrektsson T. Classification of platelet concentrates: from pure platelet-rich plasma (P-PRP) to leucocyte- and platelet-rich fibrin (L-PRF). *Trends Biotechnol* 2009;27(3):158-167.
23. Fiorellini JP, Howell TH, Cochran D et al. Randomized Study Evaluating Recombinant Human Bone Morphogenetic Protein-2 for Extraction Socket Augmentation. *J Periodontol* 2005;76:605-613.
24. Schlegel KA, Schultze-Mosgau S, Wilfang J et al. Changes of mineralization of free autogenous bone grafts used for sinus floor elevation. *Clin Oral Impl Res* 2006;17:673-678.
25. Boyan BD, Ranly DM, McMillan J et al. Osteoinductive ability of human allograft formulations. *J Periodontol* 2006;77(9):1555-63.
26. Berglundh T, Lindhe J. Healing around implants placed in bone defects treated with Bio-Oss. *Clin Oral Impl Res* 1997;8(2):117-124.
27. Artzi Z, Dayan D, Alpern Y et al. Vertical ridge augmentation using xenogenic material supported by a configured titanium mesh: clinicohistopathologic and histochemical study. *Int J Oral Maxillofac Implants* 2003;18(3):440-6.
28. Bauer TW, Muschler GF. Bone graft materials. An overview of the basic science. *Clin Orthop* 2000;371:10-27.
29. Wennerberg A, Albrektsson T. Effects of titanium surface topography on bone integration: a systematic review. *Clin Oral Implants Res* 2009;20(Suppl 4):172-84.
30. Brånemark PI, Hansson BO, Adell R et al. Osseointegrated titanium implants in the treatment of the edentulous jaw. *Scand J Plast Reconstr Surg* 1977;16:1-175.
31. Albrektsson T, Brånemark PI, Hansson HA et al. Osseointegrated titanium implants Requirements for ensuring a long-lasting, direct bone anchorage in man. *Acta Orthop Scand* 1981;52(2):155-170.
32. Riggs BL, Kholsla S, Melton LJ. A unitary model for involutional osteoporosis: estrogen deficiency caused both type I and type II osteoporosis in postmenopausal women and contributes to bone loss in aging men. *J Bone Miner Res* 1998;13:763-773.

33. Tannenbaum C, Clark J, Schwartzman K et al. Yield of laboratory testing to identify secondary contributors to osteoporosis in otherwise healthy women. *J Clin Endocrinol Metabol* 2002;87:4431-4437.
34. Albanese CV, Passariello R. *Fisiopatologia dell'osso*. In: *Osteoporosi e malattie metaboliche dell'osso*. Springer-Verlag Italia 2009.
35. Crandall C. Laboratory workup for osteoporosis Which tests are most cost-effective? *Postgrad Med* 2003;114(3):35-38,41-44.
36. Jeffcoat M. The Association Between Osteoporosis and Oral Bone Loss. *J Periodontol* 2005;76:2125-2132.
37. Esposito M, Grusovin MG, Felice P et al. Interventions for replacing missing teeth: horizontal and vertical bone augmentation techniques for dental implant treatment. *Cochrane database of systematic reviews* 2009, Issue4 Art No:CD003607.
38. Jensen SS, Hendrik Terheyden H. Bone Augmentation Procedures in Localized Defects in the Alveolar Ridge: Clinical Results with Different Bone Grafts and Bone-Substitute Materials. *Int J Oral Maxillofac Implant* 2009;24(SUPPL):218-236.
39. Hämmerle CH, Jung RE. Bone augmentation by means of barrier membranes. *Periodontology* 2000, 2003;33:36-53.
40. Maestre-Ferrin L, Boronat-López A, Peñarocha-Diago M et al. Augmentation procedures for deficient edentulous ridges, using onlay autologous grafts: An update. *Med Oral Patol Oral Cir Bucal*. 2009 Aug1;14(8):e402-7.
41. Chiapasco M, Zaniboni M, Boisco M. Augmentation procedures for the rehabilitation of deficient edentulous ridges with oral implants. *Clin Oral Impl Res* 2006;17(Suppl. 2):136-159.
42. Rawashdeh MA, Telfah H. Secondary alveolar bone grafting: the dilemma of donor site selection and morbidity. *Br J Oral Maxillofac Surg* 2008;46:665-670.
43. Hämmerle CH, Jung RE Feloutzis A. A systematic review of the survival of implants in bone sites augmented with barrier membranes (guided bone regeneration) in partially edentulous patients. *J Clin Periodontol* 2002;29(Suppl 3):226-231.
44. Aghaloo TL, Moy PK. *Wich Hard Tissue Augmentation Techniques are the most successful in furnishing Bony support for implant placement?* *Int J Oral Maxillofac Implants*. 2007,22(suppl):49-70.
45. Donors N, Mardas N, Chadha V. Clinical outcomes of implants following lateral bone augmentation: systematic assessment of available options (barrier membranes, bone grafts, split osteotomy). *J Clin Periodontol*. 2008;35(Suppl 8):173-202.
46. Rocchieta I, Fontana F, Simion M. Clinical outcomes of vertical bone augmentation to enable dental implant placement: a systematic review. *J Clin Periodontol* 2008;35(Suppl 8):203-215.
47. Fiorellini JP, Fiorellini, Nevins ML. Localize ridge augmentation/preservation: A systematic review. *Ann Periodontol* 2003;8:321-327.
48. Chiapasco M, Casentini P, Zaniboni M. Bone Augmentation Procedures in Implant Dentistry. *Int J Oral Maxillofac Implant* 2009;24(Suppl):237-259.
49. Wang HL, Boyapati L. "PASS" principles for predictable bone regeneration. *Implant Dent*. 2006;15(1):8-17.
50. Misch CE. *Densità ossea: un elemento fondamentale per il successo clinico*. In: *L'odontoiatria implantare contemporanea*. I edizione italiana. Antonio Delfino Editore 2000 pag 109-118.
51. Park SH, Brooks SL, Oh TJ, Wang HL. Effect of ridge morphology on guided bone regeneration outcome: conventional tomographic study. *J Periodontol*. 2009;80(8):1231-6.
52. Nissan J, Ghelfan O, Mardinger O et al. Efficacy of cancellous block allograft augmentation prior to implant placement in the posterior atrophic mandible. *Clin Implant Dent Related Res* 2009;5:1-7.
53. Sbordone L, Toti GB, Menchini-Fabris C et al. Volume changes of autogenous bone grafts after alveolar ridge augmentation of atrophic maxillae and mandibles. *Int J Oral Maxillofac Surg*. 2009;38:1059-1065.
54. Smolka W, Eggenesperger N, Carollo V et al. Changes in the volume and density of calvarial split bone grafts after alveolar ridge augmentation *Clin Oral Implants Res*. 2006;17(2):149-55.
55. Keith JD, Petrungraro P, Leonetti JA et al. Clinical and histological evaluation of a mineralized block allograft: result from the developmental period (2001-2004). *Int J Periodontics Rest Dent* 2006;26:321-327.
56. Ferri J, Dujoncquoy JP, Carneiro JM et al. Maxillary reconstruction to enable implant insertion: a retrospective study of 181 patients. *Head & Face Medicine* 2008, 4:31. Available at <http://www.head-face-med.com/content/4/1/31>.
57. Bahat O, Fontanessi RV. Efficacy of implant placement after bone grafting for three-dimensional reconstruction of posterior jaw. *Int J Periodontics Rest Dent* 2001;21:220-230.
58. Esposito M, Hirsch JM, Lekholm U et al. Biological factors contributing to failures of osseointegrated oral implants: (II) Etiopathogenesis. *Eur J Oral Sci* 1998;106(3):721-764.
59. Chuang SK, Wei LJ, Douglass CW et al. Risk factors for dental implant failure: a strategy for the analysis of clustered failure-time observation. *J Dent Res* 2002;81(8):572-577.
60. Hermann L, Lekholm L, Holm S et al. Evaluation of patient and implant characteristics as potential prognosis factors for oral implant failure. *Int J Oral Maxillofac Implants* 2005;20(2):220-230.
61. Trisi P, Lazzara R, Rao W, Rebaudi A. Bone-implant contact and bone quality: evaluation of expected and actual bone contact on machined and osseointegrated implant surfaces. *Int J Periodontics Restorative Dent*. 2002 Dec;22(6):535-45.
62. Crespi R, Vinci R, Cappare P et al. Calvarial versus iliac crest for autologous bone graft material for a sinus lift procedure: a histomorphometric study. *Int J Oral Maxillofac Implants* 2007;22(4):527-32.
63. Erdogan O, Shafer DM, Taxel P et al. Review of the association between osteoporosis and alveolar ridge augmentation. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2007;104:738.e1-738.e13.
64. Holahan CM, Koka S, Kennel KA et al. Effect of osteoporotic status on the survival of titanium dental implants. *Int J Oral Maxillofac Implants* 2008;23(5):905-910.
65. Cooper LF, Masuda T, Yliheikkilä PK et al. Generalizations regarding the process and phenomenon of osseointegration. Part II. In vitro studies. *Int J Oral Maxillofac Implants* 1998;13(2):163-74.
66. Narai S, Nagahata S. Effects of alendronate on the removal torque of implants in rats with induced osteoporosis. *Int J Oral Maxillofac Implants* 2003;18(2):218-23.
67. Giro G, Sakakura CE, Gonçalves D et al. Effect of 17beta-Estradiol and Alendronate on the Removal Torque of Osseointegrated Titanium Implants in Ovariectomized Rats. *J Periodontol* 2007;78:1316-1321.
68. Tokugawa Y, Shiota T, Ohno K et al. Effects of bisphosphonate on bone reaction after placement of titanium implants in tibiae of ovariectomized rats. *Int J Oral Maxillofac Implants*. 2003;18(1):66-74.
69. Tonelli P, Borgioli A, Duvina M et al. Bisphosphonate-related osteonecrosis of the jaw: the Florence experience. *Clinical Cases in Mineral and Bone Metabolism* 2007;4(1): 48-52.
70. Maïmoun L, Brennan TC, Badous I et al. Strontium ranelate improves implant osseointegration. *Bone*. 2010;46(5):1436-41.
71. Arisawa EA, Brandão AA, Almeida JD et al. Calcitonin in bone-guided regeneration of mandibles in ovariectomized rats: densitometric, histologic and histomorphometric analysis. *Int J Oral Maxillofac Surg* 2008;37(1):47-53.
72. Nociti FH Jr, Sallum AW, Sallum EA et al. Effect of estrogen replacement and calcitonin therapies on bone around titanium implants placed in ovariectomized rats: a histometric study. *Int J Oral Maxillofac Implants* 2002;17(6):786-92.
73. Anbinder AL, Junqueira JC, Mancini MN et al. Effects of simvastatin on bone regeneration in the mandibles of ovariectomized rats and on blood cholesterol levels. *J Oral Sci*. 2002;44(3-4):117-24.
74. Park JB. The use of simvastatin in bone regeneration. *Med Oral Patol Oral Cir Buccal* 2009 Sep1;14(9):e485-8.
75. Retzeppi M, Donos N. Guided Bone Regeneration: biological principle and therapeutic applications. *Clin Oral Impl Res* 2010;21:567-576.
76. Taguchi A, Sanada M, Krall E et al. Relationship between dental panoramic radiographic findings and biochemical markers of bone turnover. *J Bone Miner Res*. 2003 Sep;18(9):1689-94.
77. Drage NA, Palmer RM, Blake G et al. A comparison of bone mineral density in the spine, hip and jaws of edentulous subjects. *Clin Oral Implants Res* 2007;18(4):496-500.
78. Lee DW, Pi SH, Lee SK, Kim EC. Comparative histomorphometric analysis of extraction sockets healing implanted with bovine xenografts irradiated cancellous allografts, and solvent-dehydrated allografts in humans. *Int J Oral Maxillofac Implants*. 2009;24(4):609-15.
79. Karabunda C, Ozdemir O, Tosun T et al. Histological and clinical evaluation of 3 different grafting materials for sinus lifting procedure based on 8 cases. *J Periodontol* 2001;72:1436-1442.