Airway remodeling: the clinical significance

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Summary

While structural changes have been observed in asthmatic airways for more than a century, the importance of airway remodeling did not come into view as a key player in asthma until recently. Its significance can be derived from the fact that supramaximal doses of histamine are not capable of causing full airway constriction in healthy subjects, indicating that more structural changes must be at play. Initially it was believed that airway remodelling was caused by chronic inflammation and repeated allergen exposure resulting in a continuous process of damage and repair. Evidence is now pointing to inflammation and airway remodelling acting in parallel, perhaps mutually reinforcing each other.

The primary focus in airway remodelling research has been on airway smooth muscle changes. It is well established that airway smooth muscle mass is increased in asthma, primarily through hyperplasia, although some studies also show evidence of hypertrophy. Recent results show that airway smooths muscle proliferation in asthma, particularly severe asthma, presumably in balance with cell apoptosis, which may indicate that airway remodelling is in fact potentially reversible. It is not yet known whether airway smooth muscle force or velocity of shortening is altered in asthma, either intrinsically or modulated by the inflammatory environment of the asthmatic airway. Some in vitro and animal studies do suggest that inflammatory mediators can result in increased force generation and proliferation of airway smooth muscle.

Airway remodelling could influence airway responsiveness in a number of other ways too. An increased airway wall thickness provides a greater load for the airway smooth muscle to contract against, but also requires less airway smooth muscle shortening to achieve the same reduction in airway diameter. Recent studies have shown the bronchodilating and bronchoprotective effects of airway diameter oscillations that occur in healthy subjects during breathing. These airway diameter oscillations that are likely reduced because of airway remodelling, may abrogate the beneficial effects of breathing in asthma.

There are no known drugs that have been proven to reverse airway remodelling, although this may be partly due to the lack of accurate non-invasive tools available to measure the degree of airway remodelling. Bronchial thermoplasty, which relies on radiofrequency ablation of airway smooth muscle, has been shown to result in some improvement in quality of life in severe asthmatic patients, but minimal improvement in lung function. Identification of the dominant site of airway hyperresponsiveness may increase the effectiveness of such techniques.

KEY WORDS: bronchial asthma; airway remodelling; airway hyperresponsiveness; bronchothermoplasty.

Introduction

The link between exaggerated airway narrowing and the symptoms of asthma has been made for several centuries. The mechanistic basis for the marked bronchospasm of an acute attack of asthma has remained obscure but is generally attributed to the presence of an underlying hyperresponsiveness of the airways that can be detected by challenges with a variety of bronchoconstrictive agonists. An excessive responsiveness of the airways to stimulation with histamine was recognized in 1946 (1). However, it was not addressed as a potential consequence of altered airway smooth muscle function for several decades. Indeed there was a great deal of research that examined the possibility that control of the bronchial smooth muscle by the autonomic nervous system lay at the root of the problem. However research on the regulation of bronchomotor tone gave way to a focus on inflammatory mediators by the 1980s. From the 1990s there was renewed interest in the structural changes in the airways with the development of animal models demonstrating the plausibility of proliferation of airway smooth muscle in response to allergen challenges. Indeed the changes in muscle mass and other forms of remodeling have dominated the discourse about asthma pathogenesis. In addition the function of airway smooth muscle and its regulation have received intensive scrutiny. However if remains a great deal of uncertainty about the causes of the acute and often relatively refractory asthma attack, airway hyperresponsiveness and persistent loss of lung function associated with severe forms of asthma. In this review we will examine some of the pathophysiological processes that may help us understand these features of disease.

The neural regulation of bronchomotor tone

The airways are supplied by autonomic fibers, predominantly via the vagus. The postganglionic nerves release acetylcholine that may activate bronchial smooth muscle, and also may affect mucus gland secretion and airway blood flow (2). Studies of isolated airways demonstrate a non-cholinergic non-adrenergic negative regulatory nervous system

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that may cause bronchodilation when airway smooth muscle is contracted (3). A small number of studies have provided evidence of a similar system in vivo (4, 5). Nitric oxide seems to be one of the mediators of this relaxant system (6). There appears to be no significant direct sympathetic supply of nerves to the airway smooth muscle despite the abundance of adrenergic receptors that may mediate either contraction or relaxation. Inhibition of beta adrenergic receptors or ganglionic neurotransmission does not affect airway responsiveness to methacholine (7), indicating that the principal pathways

for neural regulation are not responsible for the enhanced airway responsiveness that is a defining feature of asthma. However, recently it has been shown that the late asthmatic response, a secondary bronchoconstrictive event which occurs 2-24 hours after the acute response, can be inhibited by anesthesia and anticholinergic drugs (8). Certain inflammatory mediators may accelerate the release of acetylcholine from post-synaptic nerve endings (9, 10). The lack of major efficacy of anti-cholinergic drugs in asthma arques against the generalized importance of such mechanisms. Although airway hyperresponsiveness may not be caused by excessive cholinergic stimulation or defective NANC relaxation, links between innervation and inflammation may have pertinence for some form of hyperalgesia that leads to cough and sputum production or indeed nerves may promote inflammation through concomitant release of neuropeptides (11). The lack of effective antagonists of neurokinins has retarded our understanding of their roles in airway disease.

Inflammatory mediators and airway hyperresponsiveness

It seemed a plausible hypothesis that an excess of inflammatory mediators could account for airway hyperresponsiveness in asthma by providing a strong stimulus to airway smooth muscle to contract. Although there is abundant evidence that inflammatory mediators trigger airway narrowing in asthma, the exaggerated narrowing seen is unlikely to

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be explained solely by this mechanism. The evidence for this statement comes from a consideration of bronchial provocation tests. Since one cannot evoke substantial falls in the forced expiratory volume in one second (FEV₁) in the majority of healthy human subjects using high concentrations of histamine (12) then an excess of other contractile agonists in the airways, such as the cysteinyl leukotrienes, should not enhance the methacholine response to the point of revealing the degree of responsiveness associated with asthma. An enhancement of the organ responsiveness is still required for excessive airway narrowing to occur. The search for inflammatory mediators that could alter airway smooth muscle properties has revealed some possible clues. Cytokines associated with asthma such as interleukin-13 and tumor necrosis factor- α enhance the contractility of airway smooth muscle (13-15) so that they may hypothetically contribute to AHR. It should be stressed that the evidence for this phenomenon has been generated in murine models and in cell culture systems. There is little or no direct evidence for this at present in human subjects. Targeting TNF- α has demonstrated little evidence for its involvement as a driver of disease and neutralization of IL-13 showed only a modest effect on lung function in asthmatics (16).

Airway remodeling and airway hyperresponsiveness

Airway remodeling or the structural changes in the airways were described by many investigators spanning over a century (17). The application of the tools of quantitative histology clearly demonstrated the increase in airway smooth muscle mass (18), subepithelial thickening (19) and a range of non-structural

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changes compared to healthy control subjects. Repeated cycles of tissue injury and repair from chronic inflammation are believed to be responsible for these structural changes, although the presence of some of the changes in childhood asthma may suggest that inflammation and remodeling occur in parallel (20, 21). Increase in airway smooth muscle mass has now been repeatedly reported (Figure 1), in particular as a function of asthma severity (22-25). It

is also associated with fixed airflow limitation in both children and adults (26, 27).



Figure 1 - Airway smooth muscle in (severe) asthma and a non-asthmatic control. Airway smooth muscle fluorescently labelled with TRITC-phalloidin in airways dissected from human lungs procured through the International Institute for Advancement of Medicine. The spiral arrangement of the airway smooth muscle is evident in both, although it appears somewhat less ordered in the severe asthma specimen.

Isolated airway smooth muscle, similar to other forms of smooth muscle, has the capacity to shorten to very short lengths. Such degrees of shortening would lead to complete closure of the airways (28). How is it then that healthy subjects do not experience marked airway closure even when challenged with high concentrations of methacholine? The answer proposed is that the force of contraction of airway smooth muscle is not sufficient to overcome the elastic resistance to deformation of the parenchyma to which the outer aspect of the airway wall is attached via alveolar attachments, as well as the intrinsic stiffness of the airway wall as it undergoes constriction (29). The effect of airway remodeling on this force balance is not fully understood. Increased airway wall thickness in remodeled airways may provide a greater load for the smooth muscle to overcome, but with the thicker airway wall the muscle needs to shorten less to achieve the same level of airway constriction (30). While the airway smooth muscle mass is increased, it is unclear whether this is associated with increases in the contractile force generated by the muscle (31-35).

Recent research has indicated that the airway diameter is not determined by a static force balance but rather a dynamic one, influenced by the constant lung volume changes that occur during normal breathing (36, 37). In fact, under static lung inflation airways do constrict to the point of airway closure (38, 39). For over half a century it has been known that deep inspirations in particular are a potent bronchodilator in healthy subjects, but not in most asthmatics (40, 41). A single deep inspiration is capable of reversing bronchoconstriction (40), while inhibition of deep inspirations results in hyperresponsiveness (42). In asthmatics deep inspiration does not lead to bronchodilation, and may even cause increased bronchoconstriction, particularly in severe asthmatics (43, 44). Tidal breathing and increased lung volume also seem to reduce bronchoconstriction from metacholine challenge in healthy subjects (45-47). In vitro studies have shown that airway smooth muscle contraction is partially inhibited when subjected to oscillatory stretches equivalent to those occurring in vivo during breathing (48). The mechanism of the "softening" of airway smooth muscle in response to stretching is still a matter of active investigation. The degree of stretch that results from a pressure applied by the expanding lung parenchyma on the airway will depend upon the stiffness of the airway wall. Fibrosis of the airway such as is described in asthma may diminish the effectiveness of breathing by reducing the length changes of the muscle caused by any given transmural pressure developed across the bronchial wall. This can be further compounded by a change in the elasticity of the parenchyma, which is characteristic of COPD, but may also occur in (severe) asthma (49).

Airway smooth muscle remodeling may not be limited to an increase in its mass. Molecular remodeling may also potentially occur, affecting the velocity of airway smooth muscle contraction. Mathematical modeling has shown that an increase in velocity of shortening of the airway smooth muscle could explain the difference in response to deep inspirations between asthmatics and controls (50). The airway smooth muscle contractile apparatus, like skeletal muscle, consists of myosin and actin filaments, with protruding myosin heads responsible for the movement of these filaments relative to each other. The 2 dominant isoforms of myosin, SM-A and SM-B, have been shown to have distinctive velocities, in motility studies, with SM-B being twice as fast as SM-A. In asthmatics the ratio of SM-B/SM-A mRNA is increased (51), but this has yet to be followed by measurements at the protein level. Alternatively a change in the ratio of myosin light chain kinase and myosin light chain phosphatase, which regulate myosin activity through phosphorylation of the myosin light chain, could result in increased shortening velocity (52).

In addition to molecular remodeling that may result in enhanced contractile properties there may be an alteration in the phenotype of the muscle. Airway smooth muscle may adopt a secretory phenotype resulting in the secretion of a variety of chemoattractants for inflammatory cells such as interleukin-8 (CXCL8; a neutrophil chemoattractant), RANTES (CCL5; a lymphocyte chemoattractant) and eotaxin (CCL11; an eosinophil chemoattractant) (53). These cells may therefore attract immune effector cells into the airways and more specifically into the muscle bundles that may then alter their properties or trigger proliferation. Proliferation of smooth muscle may be caused by contact with activated T lymphocytes (54). T cells are present in significant numbers within airway smooth muscle bundles and increase in numbers correlate with increase in severity of disease (55). There is also an increase in mast cells within the airway smooth muscle cells (56), postulated to be caused by the CXCR3/CXCL10 chemoattractant pathway (57).

Proliferation of airway smooth muscle

Morphometric measurements of airway smooth muscle mass have demonstrated a consistent increase in muscle mass in asthmatic subjects (18, 22-25). The increase in mass is linked to disease severity and not to the duration of disease (58). This suggests that host susceptibility to airway remodeling may determine the

An important issue in remodeling is whether it is fixed or a new dynamic equilibrium based on cell proliferation and cell death. outcome of environmental factors that induce asthma, such as repeated exposure to sensitizing substances. To date no studies have identified muscle-specific genes associated with asthma but it would not be surprising to discover that such associations were present.

Attempts at determining the mechanisms of the increase in muscle mass have been made by quantifying the number of nuclei per unit area of the muscle. Most studies have found a proportionate increase in nuclei to area, leading to the conclusion that proliferation of muscle or hyperplasia has occurred (59, 60). Only one study of a small number of subjects with severe asthma using a marker of cellular proliferation, proliferating cell nuclear antigen, has identified an excess rate of proliferation of muscle (61). The authors have concluded that muscle remodeling is not only through proliferation of muscle cells but is a dynamic phenomenon. If confirmed, and this study requires confirmation, then it would imply that there is a balance between cellular apoptosis and proliferation that sets a new balance of increased muscle mass in these subjects. It also implies that muscle remodeling is not necessarily an irreversible phenomenon.

Some studies have also found evidence of increased mass that is attributable to hypertrophy (24, 62). The pattern of hypertrophy, whether associated with small or large airways also seems to vary. Some subjects demonstrate hypertrophy in large airways but not in small, whereas others have no evidence for hypertrophy. The significance of these changes (Figure 2) for airway function is unknown, although the contractility of hypertrophic muscle is likely to be impaired compared to normal muscle or muscle that has undergone an increase through hyperplasia. Understanding the implications of these different patterns of growth of muscle will require appropriate measurements of the mechanical properties of these muscles and perhaps also the elucidation of the mechanisms of the changes. Many studies have shown the plausibility of smooth muscle growth through the actions of various mediators present in the inflamed airways (63). An array of substances from histamine to chemokines such as IL-8 and classical growth factor receptors such as the epidermal growth factor receptor (EGFR) has been implicated in the growth of smooth muscle. Exploration of airway smooth muscle growth has been largely dependent on models of asthma. Both rat and mouse models have been developed based on allergic sensitization and subsequent repeated exposures to the allergen. Anti-leukotrienes and inhibitors of the epidermal growth factor receptor have been demonstrated to inhibit smooth muscle growth (64, 65). Not surprisingly compounds that are effective in inhibiting some component of the allergic response also inhibit smooth muscle growth. Thus it has not been clearly established to what extent the inflammatory process is necessary for smooth muscle growth. Mechanical stress may release EGFR ligands (66) and lead to remodeling without inflammation, raising the possibility that the very process of bronchoconstriction could lead to remodeling. Anti-cholinergic drugs also inhibit remodeling but the mechanism may go beyond inhibition of airway narrowing (67, 68). Less is known about the process of hypertrophy. In vitro studies suggest that transforming growth factor- β may cause upregulation of smooth muscle contractile proteins (69). This mediator has many actions and is likely present in asthma. The consequences of hypertrophy of airway smooth muscle require further exploration.

An important issue in remodeling is whether it is fixed or a new dynamic equilibrium based on cell proliferation and cell death. Normal tissue remodeling is a function of cellular turnover and although smooth muscle is more slowly proliferating than epithelium, for example there is evidence of some proliferation of cells in normal animals, even large mammals such as the horse (70). There is less information available for human subjects but one study has suggested that at least in severe asthma airway smooth muscle may be in a state of dynamic remodeling (61) and therefore amenable to therapy.



Figure 2 - Schematic drawing of features of airway remodelling influencing AHR. Remodelled and control airway side-by-side comparison. 1) Increased collagen content may lead to stiffening of the airway wall, providing a greater load against which the airway smooth muscle must shorten. 2) Basement membrane folding, caused by the incompressibility of the airway wall, results in a dramatic decrease of the open airway diameter, and trapping of mucous. However, basement membrane folding may also provide an increased load against airway smooth muscle shortening. 3) Airway smooth muscle hyperplasia may increase force generating potential of the airway smooth muscle, but not all cells may be in an optimal contractile state. 4) Parenchymal tethering may be reduced in asthma, but little evidence of this has been found. 5) Airway wall thickening inwards from the airway smooth muscle layer enhances airway constriction, while outwards thickening increases the resistive load to airway constriction.

Therapeutic approaches

To date there are no known drugs proven to reverse airway smooth muscle remodeling, which may in part be due to the lack of methods to non-invasively assess the degree of remodeling. It may be possible that inhibition of airway inflammation has a beneficial effect on remodeling, but it will be necessary to assess muscle remodeling more directly in order to draw conclusions about this. Newer imaging techniques such as optical

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coherence tomography may provide information of this nature but thus far its usefulness has not been established. CT imaging has proven to be unhelpful in assessing the degree of remodeling in some studies of severe asthma and is a technique that may provide information about airway wall thickness (71). Resolving remodeling from bronchoconstriction may be more

difficult to achieve. In severe asthma with fixed airflow limitation and biopsy-proven airway smooth muscle thickening, CT scans did not detect the abnormality (27).

Bronchothermoplasty is the only current treatment that specifically targets airway smooth muscle with a view to its destruction (72). Radiofrequency ablation is attempted at three bronchoscopic sessions. The effectiveness of the intervention is limited to improvements in symptoms and in rates of exacerbation (73). Substantial change in lung function is not reported. Confirmation of the removal of smooth muscle is supported by human and animal studies (74, 75). It seems reasonable that this postulated mechanism of action does indeed account for clinical outcomes. One could imagine that incomplete ablation could limit improvement since airway narrowing could still occur in inadequately treated areas. Further refinements in this direction seem a promising approach to curing asthma. Many subjects will likely not benefit from thermoplasty if the site of their airway remodeling is distal to the generations of airways targeted by treatment. Identification of the dominant site of airway smooth muscle remodeling would permit appropriate selection of subjects and appropriate targeting of airways.

Conclusion

The search for the culprit in airway hyperresponsiveness has changed focus several times over the years. Because of our limited knowledge of the precise significance of airway remodeling the therapeutic approaches to limit or reverse it are uncertain. While the research to date has given us some indication on what is *not* causing airway hyperresponsiveness, the explanation for the observed difference between the apparent behaviour of airway smooth muscle *in vivo* in asthmatic and healthy subjects remains

elusive. Airway remodeling certainly plays an important role in AHR, but the jury is still out on its causes and effects. Are injury and repair responsible or are there innate differences in at least some asthmatics, as suggested by findings in childhood asthma? Does airway remodeling provide an element of bronchoprotection through increased airway wall stiffness, or is increasing AHR through increased muscle mass and subepithelial thickness?

Because of our limited knowledge of the precise significance of airway remodeling the therapeutic approaches to limit or reverse it are uncertain. Novel approaches to the ablation of airway smooth muscle through bronchial thermoplasty await more extensive critical studies of efficacy and the optimal choice of subject. Will those with excess smooth muscle prove to be the most benefitted? Perhaps better localization of the precise site of AHR will assist in determining the best approaches to therapy.

References

- 1. Curry JJ. The action of histamine on the respiratory tract in normal and asthmatic subjects. J Clin Invest 1946;25(6):785-791.
- 2. Barnes PJ. Overview of neural mechanisms in asthma. Pulm Pharmacol 1995;8(4-5):151-159.
- Li CG, Rand MJ. Evidence that part of the NANC relaxant response of guinea-pig trachea to electrical field stimulation is mediated by nitric oxide. Br J Pharmacol 1991;102:91-94.
- Diamond L, O'Donnell M. A nonadrenergic vagal inhibitory pathway to feline airways. Science 1980;208(4440):185-188.
- Irvin CG, Boileau R, Tremblay J, Martin RR, Macklem PT. Bronchodilatation: noncholinergic, nonadrenergic mediation demonstrated in vivo in the cat. Science 1980;207(4432):791-792.
- Belvisi MG, Stretton CD, Yacoub M, Barnes PJ. Nitric oxide is the endogenous neurotransmitter of bronchodilator nerves in humans. Eur J Pharmacol 1992;210:221-222.
- Sterk PJ, Daniel EE, Zamel N, Hargreave FE. Limited maximal airway narrowing in nonasthmatic subjects. Role of neural control and prostaglandin release. Am Rev Respir Dis 1985;132:865-870.
- Raemdonck K, de Alba J, Birrell MA, Grace M, Maher SA, Irvin CG, Fozard JR, O'Byrne PM, Belvisi MG. A role for sensory nerves in the late asthmatic response. Thorax 2012;67(1):19-25.
- 9. Shore S, Irvin CG, Shenkier T, Martin JG. Mechanisms of histamine-induced contraction of canine

airway smooth muscle. J Appl Physiol Respir Environ Exerc Physiol 1983;55:22-26.

- Hall AK, Barnes PJ, Meldrum LA, Maclagan J. Facilitation by tachykinins of neurotransmission in guineapig pulmonary parasympathetic nerves. Br J Pharmacol 1989;97(1):274-280.
- Vergnolle N, Cenac N, Altier C, Cellars L, Chapman K, Zamponi GW, Materazzi S, Nassini R, Liedtke W, Cattaruzza F, Grady EF, Geppetti P, Bunnett NW. A role for transient receptor potential vanilloid 4 in tonicity-induced neurogenic inflammation. Br J Pharmacol 2010;159(5):1161-1173.
- Cockcroft DW, Killian DN, Mellon JJ, Hargreave FE. Bronchial reactivity to inhaled histamine: a method and clinical survey. Clin Allergy 1977;7:235-243.
- Deshpande DA, Dogan S, Walseth TF, Miller SM, Amrani Y, Panettieri RA, Kannan MS. Modulation of calcium signaling by interleukin-13 in human airway smooth muscle: role of CD38/cyclic adenosine diphosphate ribose pathway. Am J Respir Cell Mol Biol 2004;31(1):36-42.
- Risse PA, Jo T, Suarez F, Hirota N, Tolloczko B, Ferraro P, Grutter P, Martin JG. Interleukin-13 inhibits proliferation and enhances contractility of human airway smooth muscle cells without change in contractile phenotype. Am J Physiol Lung Cell Mol Physiol 2011;300(6):L958-L966.
- Shore SA, Moore PE. Effects of cytokines on contractile and dilator responses of airway smooth muscle. Clinical and Experimental Pharmacology and Physiology 2002;29(10):859-866.
- Corren J, Lemanske RF, Hanania NA, Korenblat PE, Parsey MV, Arron JR, Harris JM, Scheerens H, Wu LC, Su Z, Mosesova S, Eisner MD, Bohen SP, Matthews JG. Lebrikizumab treatment in adults with asthma. N Engl J Med 2011;365(12):1088-1098.
- Huber HL, Koessler KK. The pathology of bronchial asthma. Archives of Internal Medicine 1922;30:689-760.
- Dunnill MS, Masarrella GR, Anderson JA. A comparison of the quantitative anatomy of the bronchi in normal subjects, in status asthmaticus, in chronic bronchitis and in emphysema. Thorax 1969;24:176-179.
- Chetta A, Foresi A, Del Donno M, Bertorelli G, Pesci A, Olivieri D. Alrways remodeling is a distinctive feature of asthma and is related to severity of disease. CHEST Journal 1997;111(4):852-857.
- Barbato A, Turato G, Baraldo S, Bazzan E, Calabrese F, Panizzolo C, Zanin ME, Zuin R, Maestrelli P, Fabbri LM, Saetta M. Epithelial Damage and Angiogenesis in the Airways of Children with Asthma. American Journal of Respiratory and Critical Care Medicine 2006;174(9):975-981.
- Payne DNR, Rogers AV, Ädelroth E, Bandi V, Guntupalli KK, Bush A, Jeffery PK. Early Thickening of the Reticular Basement Membrane in Children with Difficult Asthma. American Journal of Respiratory and Critical Care Medicine 2003;167(1):78-82.
- 22. Kuwano K, Bosken CH, Pare PD, Bai TR, Wiggs BR, Hogg JC. Small airways dimensions in asthma and in chronic obstructive pulmonary disease. Am Rev Respir Dis 1993;148:1220-1225.

- Pepe C, Foley S, Shannon J, Lemiere C, Olivenstein R, Ernst P, Ludwig MS, Martin JG, Hamid Q. Differences in airway remodeling between subjects with severe and moderate asthma. J Allergy Clin Immunol 2005;116(3):544-549.
- James AL, Elliot JG, Jones RL, Carroll ML, Mauad T, Bai TR, Abramson MJ, Mckay KO, Green FH. Airway smooth muscle hypertrophy and hyperplasia in asthma. Am J Respir Crit Care Med 2012;185(10): 1058-1064.
- Benayoun L, Druilhe A, Dombret MC, Aubier M, Pretolani M. Airway structural alterations selectively associated with severe asthma. Am J Respir Crit Care Med 2003;167(10):1360-1368.
- 26. Tillie-Leblond I, de Blic J, Jaubert F, Wallaert B, Scheinmann P, Gosset P. Airway remodeling is correlated with obstruction in children with severe asthma. Allergy 2008;63(5):533-541.
- Kaminska M, Foley S, Maghni K, Storness-Bliss C, Coxson H, Ghezzo H, Lemiere C, Olivenstein R, Ernst P, Hamid Q, Martin J. Airway remodeling in subjects with severe asthma with or without chronic persistent airflow obstruction. J Allergy Clin Immunol 2009;124(1):45-51.
- Macklem PT. A theoretical analysis of the effect of airway smooth muscle load on airway narrowing. Am J Respir Crit Care Med 1996;153(1):83-89.
- 29. Lambert RK, Wiggs BR, Kuwano K, Hogg JC, Pare PD. Functional significance of increased airway smooth muscle in asthma and COPD. J Appl Physiol 1993;74(6):2771-2781.
- Lambert RK, Wiggs BR, Kuwano K, Hogg JC, P.D. Pare PD. Functional significance of increased airway smooth muscle in asthma and COPD. J Appl Physiol 1993;74(6):2771-2781.
- Bai TR. Abnormalities in Airway Smooth Muscle in Fatal Asthma: A Comparison between Trachea and Bronchus. American Journal of Respiratory and Critical Care Medicine 1991;143(2):441-443.
- Bai TR, w.t.t.a.o.F.W. Prasad. Abnormalities in Airway Smooth Muscle in Fatal Asthma. American Journal of Respiratory and Critical Care Medicine 1990;141(3): 552-557.
- Goldie RG, Spina D, Henry PJ, Lulich KM, Paterson JW. In vitro responsiveness of human asthmatic bronchus to carbachol, histamine, beta-adrenoceptor agonists and theophylline. British Journal of Clinical Pharmacology 1986;22(6):669-676.
- Whicker SD, Armour CL, Black JL. Responsiveness of bronchial smooth muscle from asthmatic patients to relaxant and contractile agonists. Pulmonary Pharmacology 1988;1(1):25-31.
- Chin LYM, Bossé Y, Pascoe C, Hackett TL, Seow CY, Paré PD. Mechanical properties of asthmatic airway smooth muscle. European Respiratory Journal 2012;40(1):45-54.
- Bates JHT, Lauzon A.-M. Parenchymal tethering, airway wall stiffness, and the dynamics of bronchoconstriction. J Appl Physiol 2007;102(5):1912-1920.
- 37. Shen X, Gunst SJ, Tepper RS. Effect of tidal volume and frequency on airway responsiveness in mechan-

ically ventilated rabbits. J Appl Physiol 1997;83(4): 1202-1208.

- Warner DO, Gunst SJ. Limitation of Maximal Bronchoconstriction in Living Dogs. American Review of Respiratory Disease 1992;145(3):553-560.
- Gunst SJ, Warner DO, Wilson TA, Hyatt RE. Parenchymal interdependence and airway response to methacholine in excised dog lobes. J Appl Physiol 1988;65(6):2490-2497.
- Nadel JA, Tierney DF. Effect of a previous deep inspiration on airway resistance in man. J Appl Physiol 1961;16(4):717-719.
- Fish JE, Peterman VI, Cugell DW. Effect of deep inspiration on airway conductance in subjects with allergic rhinitis and allergic asthma. Journal of Allergy and Clinical Immunology 1977;60(1):41-46.
- Kapsali T, Permutt S, Laube B, Scichilone N, Togias A. Potent bronchoprotective effect of deep inspiration and its absence in asthma. J Appl Physiol 2000;89 (2):711-720.
- 43. Gayrard P, Orehek J, Grimaud C, Charpin J. Bronchoconstrictor effects of a deep inspiration in patients with asthma. The American review of respiratory disease 1975;111(4):433-439.
- 44. Salome CM, Thorpe CW, Diba C, Brown NJ, Berend N, King GG. Airway re-narrowing following deep inspiration in asthmatic and nonasthmatic subjects. European Respiratory Journal 2003;22(1):62-68.
- Irvin CG. Lung volume: a principle determinant of airway smooth muscle function. Eur Respir J 2003;22 (1):3-5.
- Salerno FG, Shinozuka N, Fredberg JJ, Ludwig MS. Tidal volume amplitude affects the degree of induced bronchoconstriction in dogs. J Appl Physiol 1999;87 (5):1674-1677.
- Shen X, Gunst SJ, Tepper R. Effect of tidal volume and frequency on airway responsiveness in mechanically ventilated rabbits. J Appl Physiol 1997;83(4): 1202-1208.
- Fredberg JJ, Inouye D, Miller B, Nathan M, Jafari S, Raboudi SH, Butler JP, Shore SA. Airway smooth muscle, tidal stretches, and dynamically determined contractile states. Am J Respir Crit Care Med 1997;156(6):1752-1759.
- Kraft M, Djukanovic R, Wilson S, Holgate ST, Martin RJ. Alveolar tissue inflammation in asthma. American Journal of Respiratory and Critical Care Medicine 1996;154(5):1505-1510.
- Bullimore SR, Siddiqui S, Donovan GM, Martin JG, Sneyd J, Bates JHT, Lauzon A.-M. Could an increase in airway smooth muscle shortening velocity cause airway hyperresponsiveness? Am J Physiol Lung Cell Mol Physiol 2011;300(1):L121-L131.
- Léguillette R, Laviolette M, Bergeron C, Zitouni N, Kogut P, Solway J, Kachmar L, Hamid Q, Lauzon A.-M. Myosin, Transgelin, and Myosin Light Chain Kinase. American Journal of Respiratory and Critical Care Medicine 2009;179(3):194-204.
- Ijpma G, Matusovsky O, Lauzon A.-M. Accumulating Evidence for Increased Velocity of Airway Smooth Muscle Shortening in Asthmatic Airway Hyperresponsiveness. Journal of Allergy 2012. 2012:5.

- 53. Damera G, Panettieri Jr RA. Does airway smooth muscle express an inflammatory phenotype in asthma? Br J Pharmacol 2011;163(1):68-80.
- Ramos-Barbon D, Presley JF, Hamid QA, Fixman ED, Martin JG. Antigen-specific CD4+ T cells drive airway smooth muscle remodeling in experimental asthma. J Clin Invest 2005;115(6):1580-1589.
- 55. Ramos-Barbon D, Fraga-Iriso R, Brienza NS, Montero-Martinez C, Verea-Hernando H, Olivenstein R, Lemiere C, Ernst P, Hamid QA, Martin JG. T Cells Localize with Proliferating Smooth Muscle {alpha}-Actin+ Cell Compartments in Asthma. Am J Respir Crit Care Med 2010.
- Brightling CE, Bradding P, Symon FA, Holgate ST, Wardlaw AJ, Pavord ID. Mast-cell infiltration of airway smooth muscle in asthma. N Engl J Med 2002;346 (22):1699-1705.
- 57. Brightling CE, Ammit AJ, Kaur D, Black JL, Wardlaw AJ, Hughes JM, Bradding P. The CXCL10/CXCR3 axis mediates human lung mast cell migration to asthmatic airway smooth muscle. Am J Respir Crit Care Med 2005;171(10):1103-1108.
- James AL, Bai TR, Mauad T, Abramson MJ, Dolhnikoff M, Mckay KO, Maxwell PS, Elliot JG, Green FH. Airway smooth muscle thickness in asthma is related to severity but not duration of asthma. Eur Respir J 2009;34(5):1040-1045.
- 59. Heard BE, Hossain S. Hyperplasia of bronchial muscle in asthma. J Pathol 1973;110:319-331.
- Woodruff PG, Dolganov GM, R.E. Ferrando RE, Donnelly S, Hays SR, Solberg OD, Carter R, Wong HH, Cadbury PS, Fahy JV. Hyperplasia of smooth muscle in mild to moderate asthma without changes in cell size or gene expression. Am J Respir Crit Care Med 2004;169(9):1001-1006.
- Hassan M, Jo T, Risse PA, Tolloczko B, Lemiere C, Olivenstein R, Hamid Q, Martin JG. Airway smooth muscle remodeling is a dynamic process in severe long-standing asthma. J Allergy Clin Immunol 2010;125(5):1037-1045.
- Ebina M, Takahashi T, Chiba T, Motomiya M. Cellular hypertrophy and hyperplasia of airway smooth muscles underlying bronchial asthma. A 3-D morphometric study. Am Rev Respir Dis 1993;148(3):720-726.
- 63. Hirota N, Martin JG. Mechanisms of airway remodeling. Chest 2013;144(3):1026-1032.
- 64. Henderson WR, Tang LO, Chu SJ, Tsao SM, Chiang GKS, Jones F, Jonas M, Pae C, Wang HJ, Chi EY. A role for cysteinyl leukotrienes in airway remodeling in a mouse asthma model. Am J Respir Crit Care Med 2002;165(1):108-116.
- 65. Tamaoka M, Hassan M, McGovern T, Ramos-Barbon D, Jo T, Yoshizawa Y, Tolloczko B, Hamid Q, Martin

JG. The epidermal growth factor receptor mediates allergic airway remodelling in the rat. Eur Respir J 2008;32(5):1213-1223.

- Tschumperlin DJ, Dai G, Maly IV, Kikuchi T, Laiho LH, McVittie AK, Haley KJ, Lilly CM, So PT, Lauffenburger DA, Kamm RD, Drazen JM. Mechanotransduction through growth-factor shedding into the extracellular space. Nature 2004;429(6987):83-86.
- Gosens R, Bos IS, Zaagsma J, Meurs H. Protective effects of tiotropium bromide in the progression of airway smooth muscle remodeling. Am J Respir Crit Care Med 2005;171(10):1096-1102.
- Bos IS, Gosens R, Zuidhof AB, Schaafsma D, Halayko AJ, Meurs H, Zaagsma J. Inhibition of allergeninduced airway remodelling by tiotropium and budesonide: a comparison. Eur Respir J 2007;30(4): 653-661.
- Goldsmith AM, Bentley JK, Zhou L, Jia Y, Bitar KN, Fingar DC, Hershenson MB. Transforming growth factor-beta induces airway smooth muscle hypertrophy. Am J Respir Cell Mol Biol 2006;34(2):247-254.
- Leclere M, Lavoie-Lamoureux A, Joubert P, Relave F, Lanctot SE, Beauchamp G, Couture C, Martin JG, Lavoie JP. Corticosteroids and Antigen Avoidance Decrease Airway Smooth Muscle Mass in an Equine Asthma Model. Am J Respir Cell Mol Biol 2012.
- Shimizu K, Hasegawa M, Makita H, Nasuhara Y, Konno S, Nishimura M. Comparison of airway remodelling assessed by computed tomography in asthma and COPD. Respiratory Medicine 2011;105 (9):1275-1283.
- Cox G, Miller JD, McWilliams A, FitzGerald JM, Lam S. Bronchial thermoplasty for asthma. Am J Respir Crit Care Med 2006;173(9):965-969.
- 73. Castro M, Rubin AS, Laviolette M, Fiterman J, De Andrade LM, Shah PL, Fiss E, Olivenstein R, Thomson NC, Niven RM, Pavord ID, Simoff M, Duhamel DR, McEvoy C, Barbers R, Ten Hacken NH, Wechsler ME, Holmes M, Phillips MJ, Erzurum S, Lunn W, Israel E, Jarjour N, Kraft M, Shargill NS, Quiring J, Berry SM, Cox G. Effectiveness and safety of bronchial thermoplasty in the treatment of severe asthma: a multicenter, randomized, double-blind, sham-controlled clinical trial. Am J Respir Crit Care Med 2010;181(2):116-124.
- Miller JD, Cox G, Vincic L, Lombard CM, Loomas BE, Danek CJ. A prospective feasibility study of bronchial thermoplasty in the human airway. Chest 2005;127 (6):1999-2006.
- 75. Brown RH, Wizeman W, Danek C, Mitzner W. In vivo evaluation of the effectiveness of bronchial thermoplasty with computed tomography. J Appl Physiol (1985) 2005;98(5):1603-1606.